

The Yeast Deletion Collection: 6000 Strains

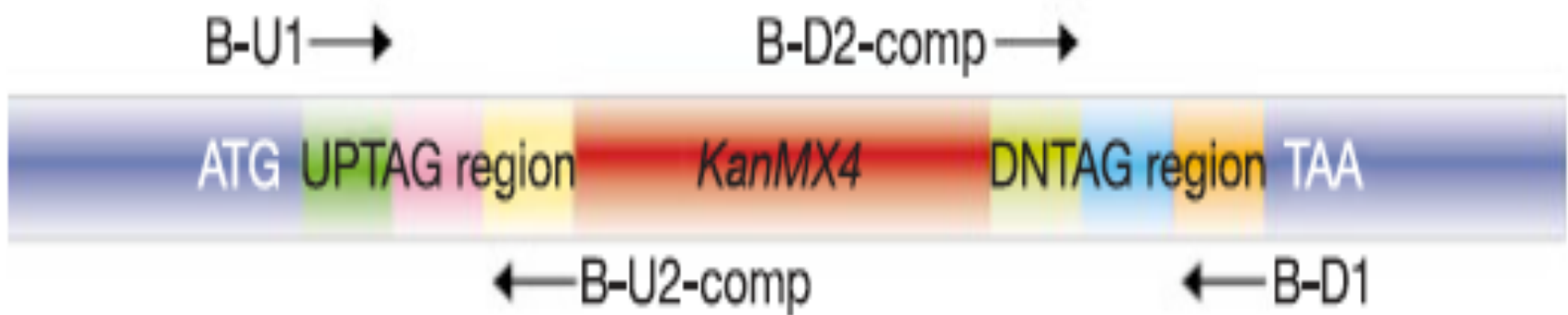
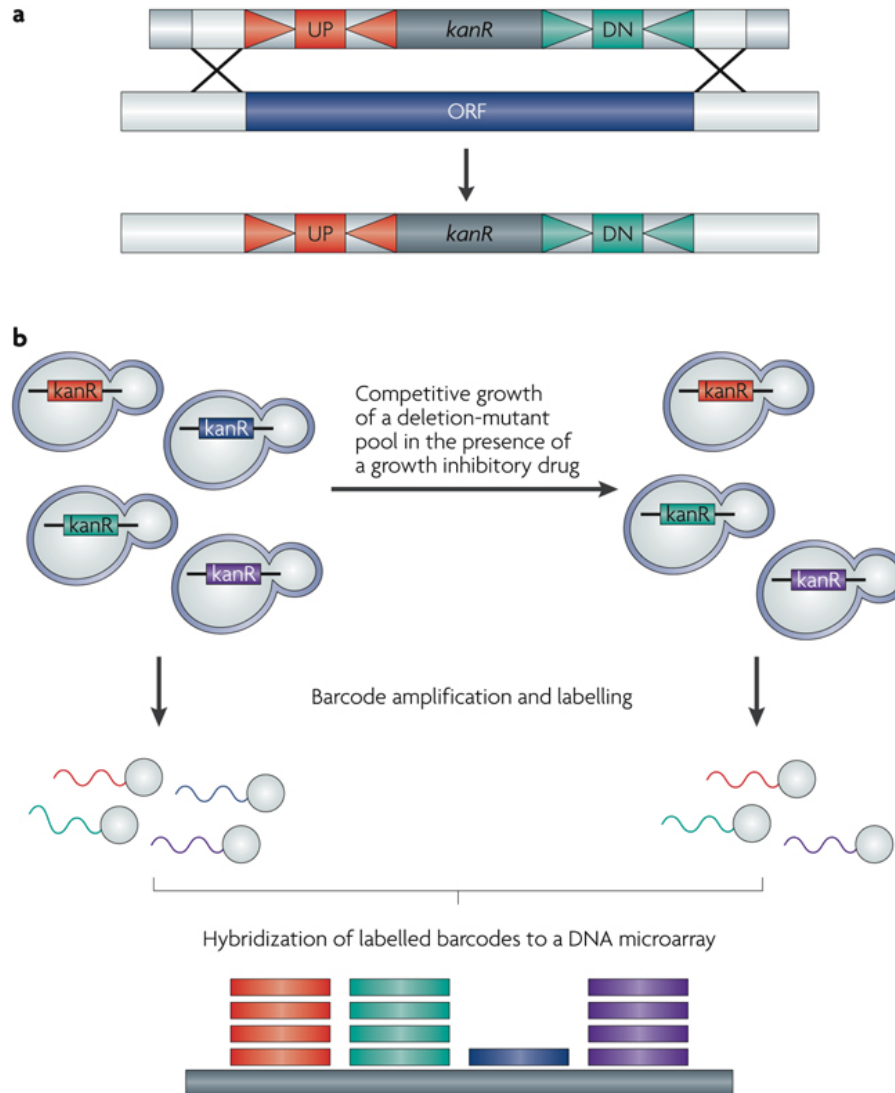
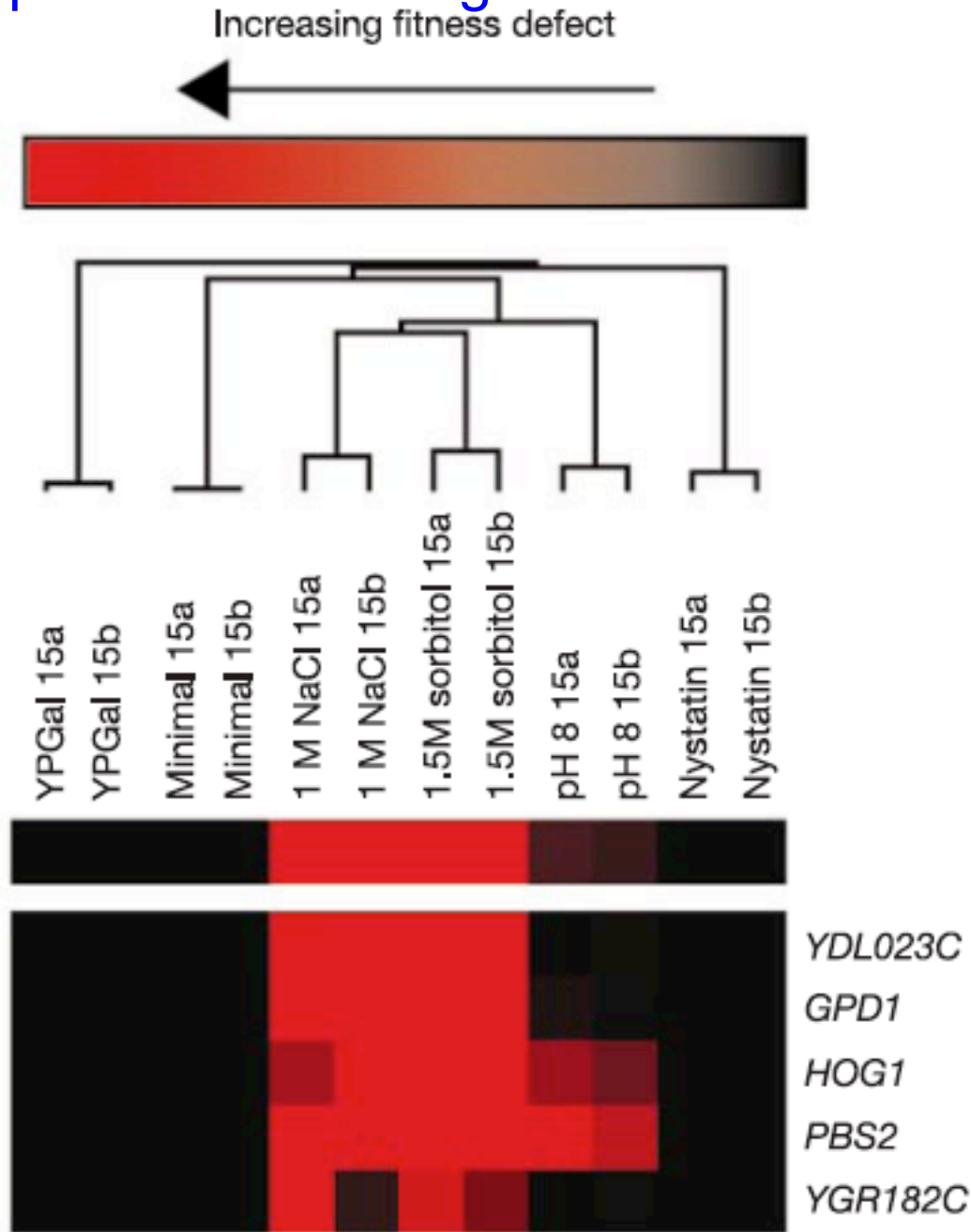


Figure 1 The *KanMX* deletion cassette module. The biotin-labelled, deletion-specific primers (B-U1, B-U2-comp, B-D1 and B-D2-comp; see Methods for structure) are used to amplify the unique UPTAG and DNTAG sequences from genomic preparations generated in the fitness-profiling studies.

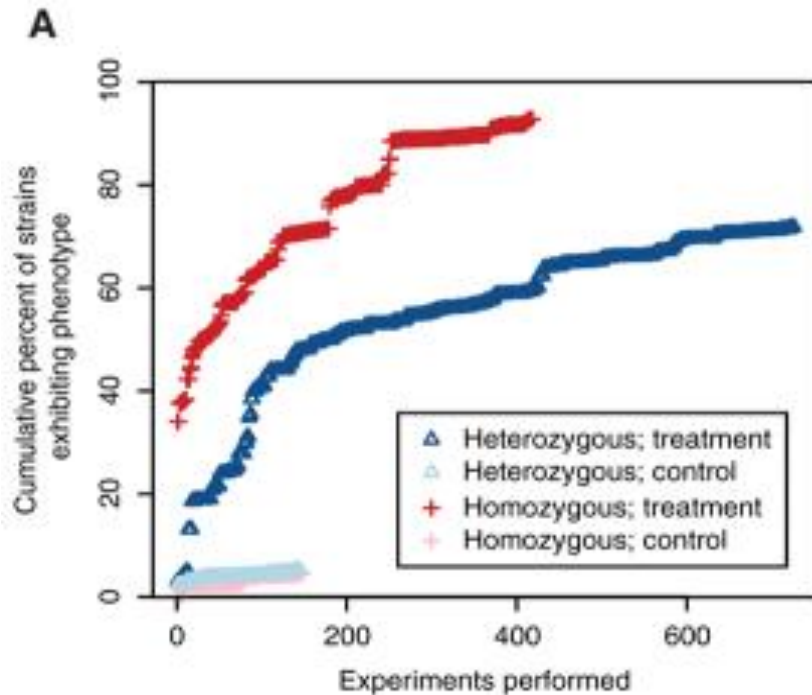
Competitive Fitness Tests



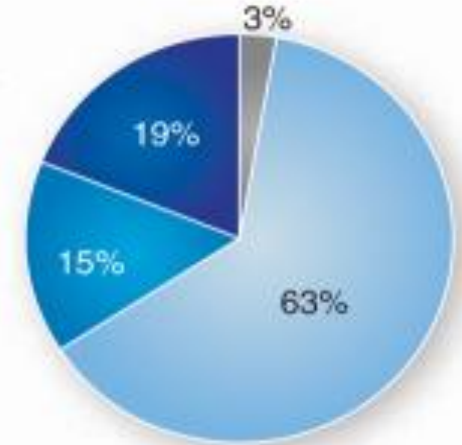
A small sample of tests: new genes sensitive to salt/sorbitol



Chemical genomic profiling with ~700 chemicals reveals a phenotype for virtually every gene deletion



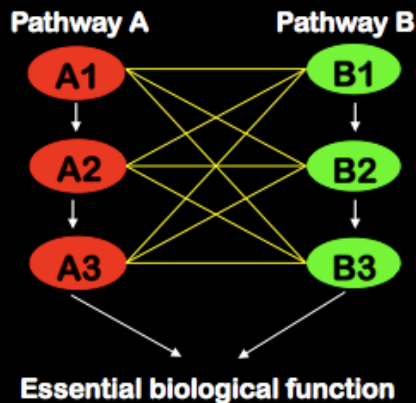
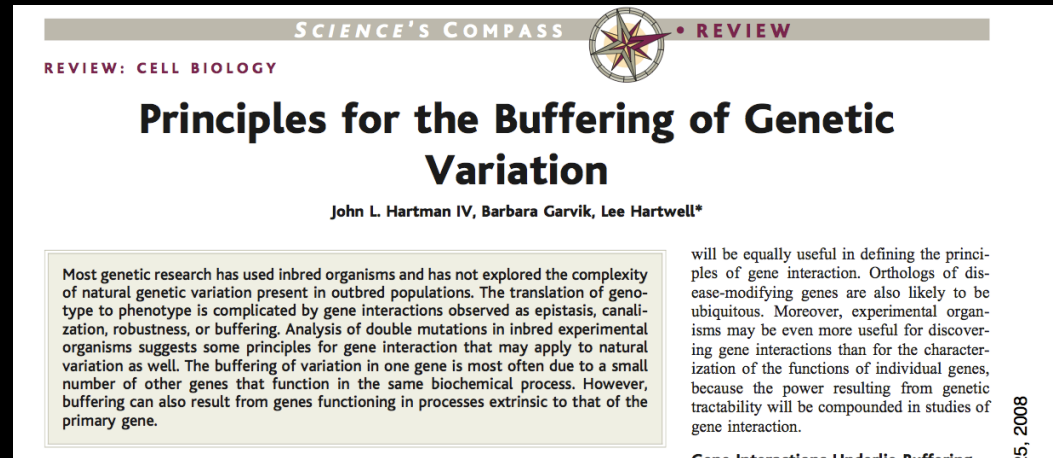
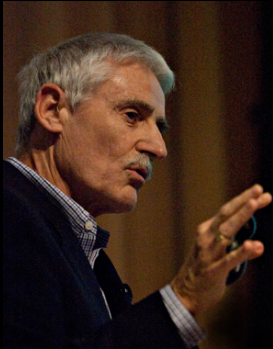
B *Consequences of deletions:*



Published in 2008, so Mark Johnston's prediction wasn't far wrong
... depending on your definition of "know"

The Hartwell Idea

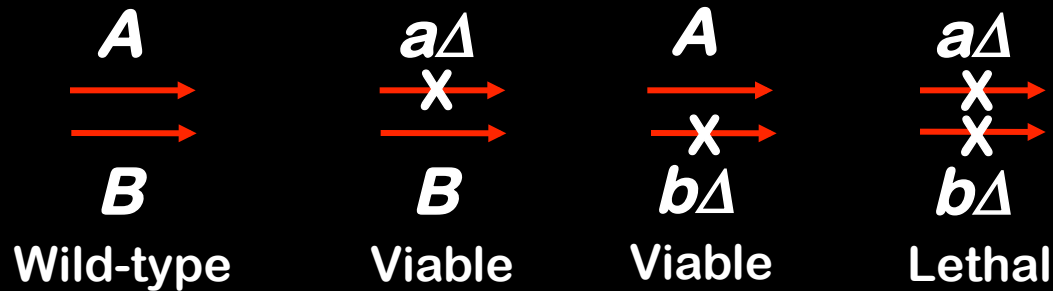
Synthetic Lethal Networks May Guide Our Understanding of Genotype to Phenotype



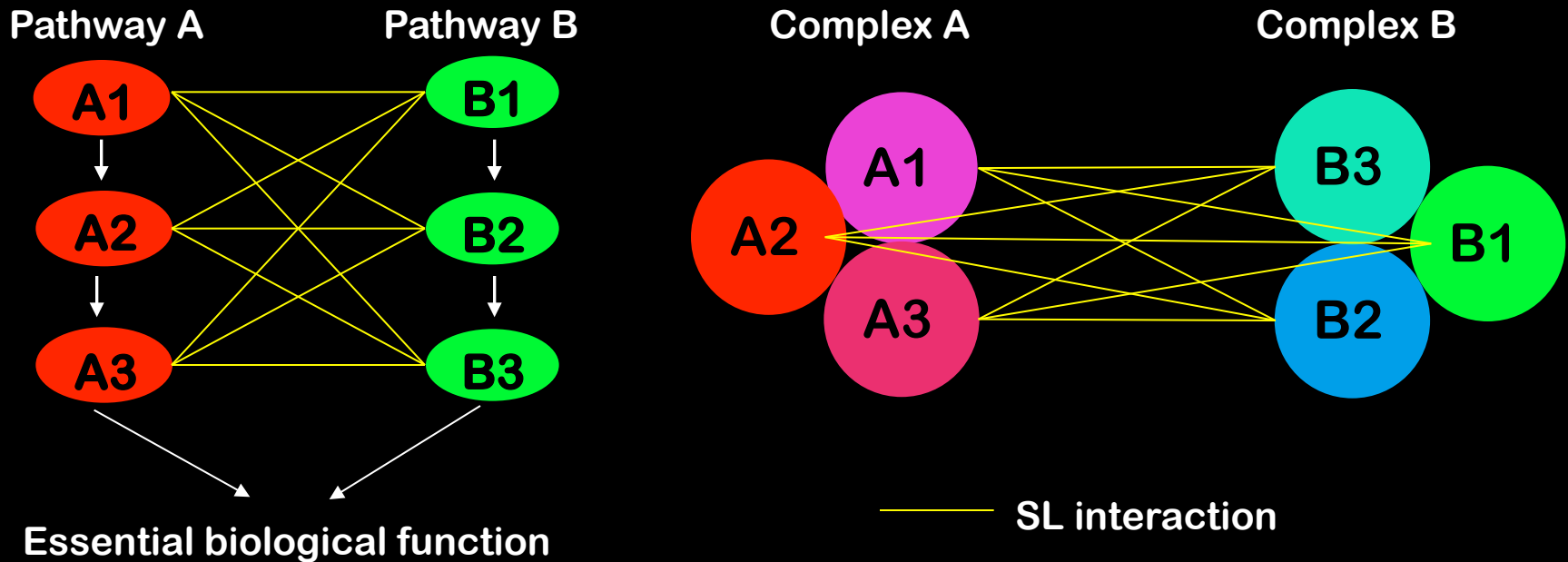
Hartman, Garvik, and Hartwell,
Science, 9 February (2001), Pg. 1001

“analysis of double mutations in inbred experimental organisms suggest principles that may apply to natural variation in outbred populations”

Synthetic Lethality



Functional Relationships



Multiplicative Model

Expected Double Mutant Fitness

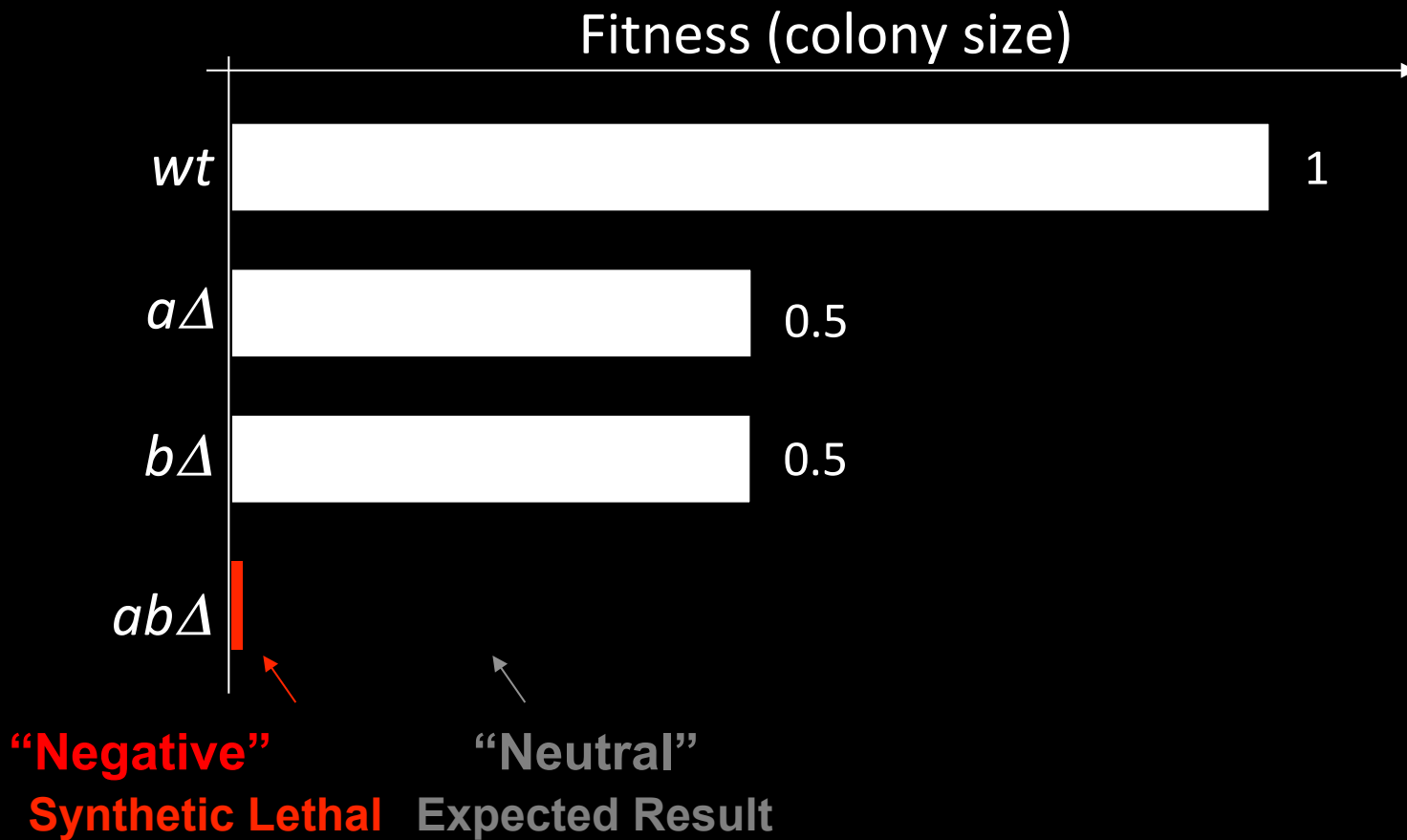


D. Segre, R. Kishony et al., *Nature Genetics* **37**, 77 - 83 (2004)

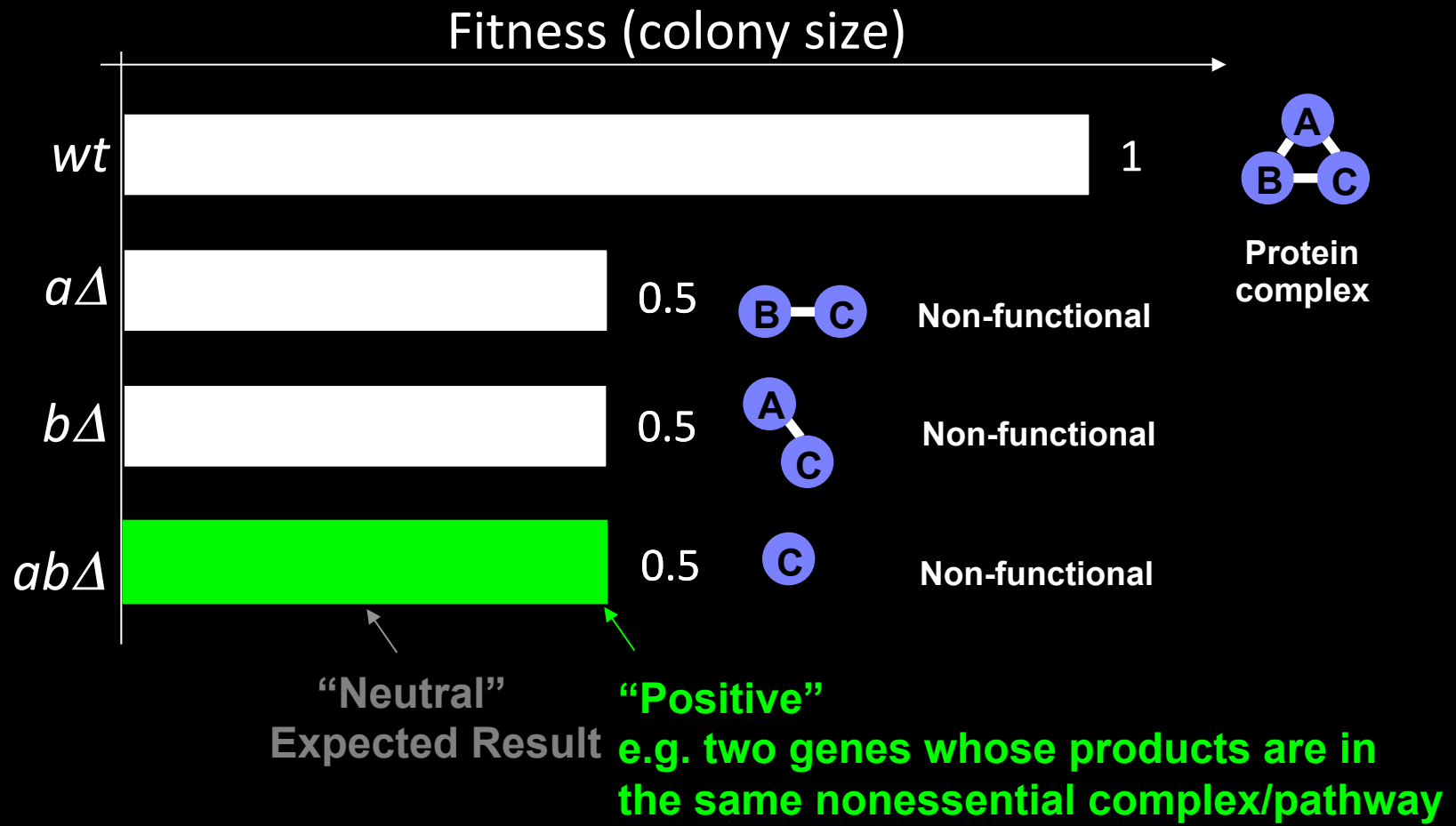
S. R. Collins, N. Krogan, J. Weissman, *Genome Biol* 2006;7:R63. “S-Score”

R. Mani F. Roth et. al., *PNAS* 2008 Mar 4;105(9):3461-6. Epub 2008 Feb 27

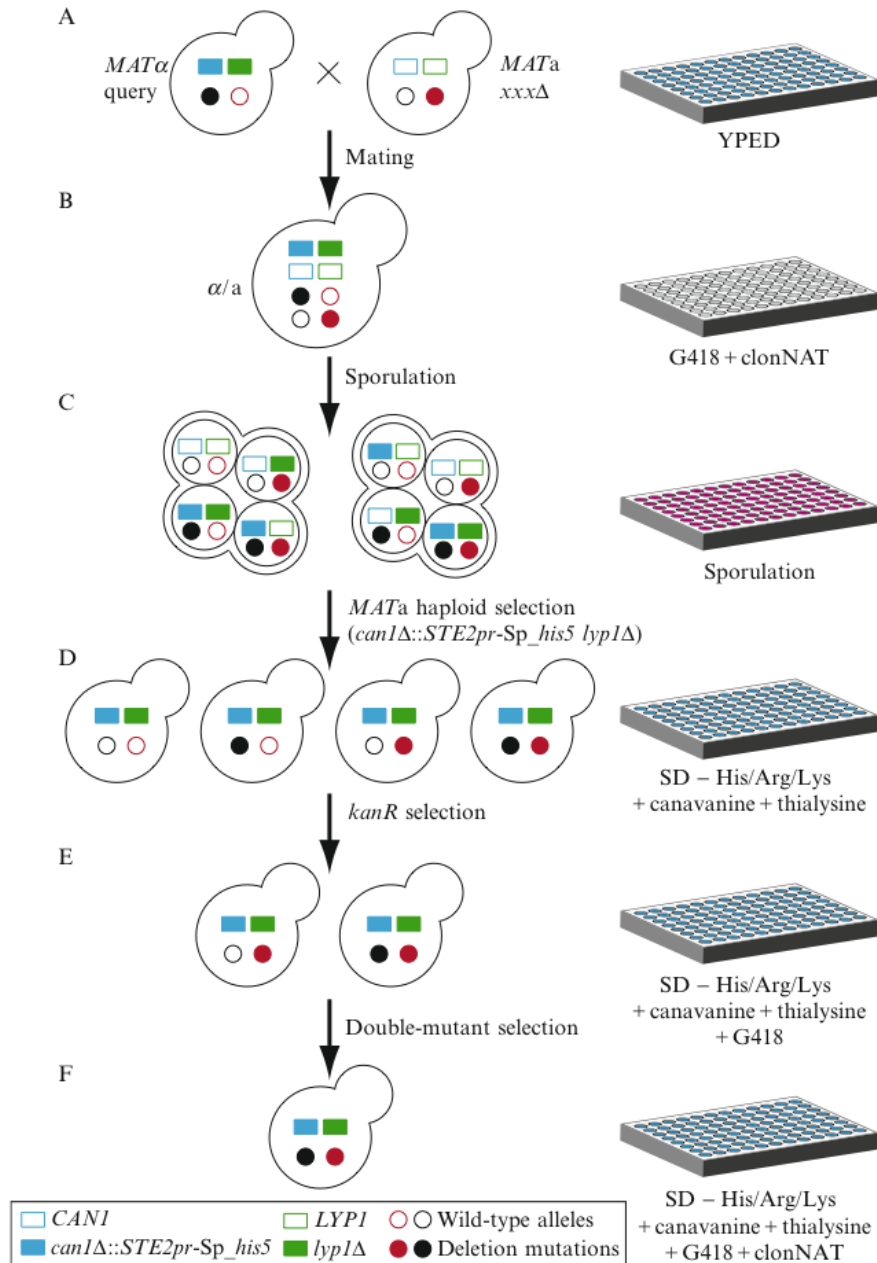
Two Basic Types of Genetic Interactions



Two Basic Types of Genetic Interactions

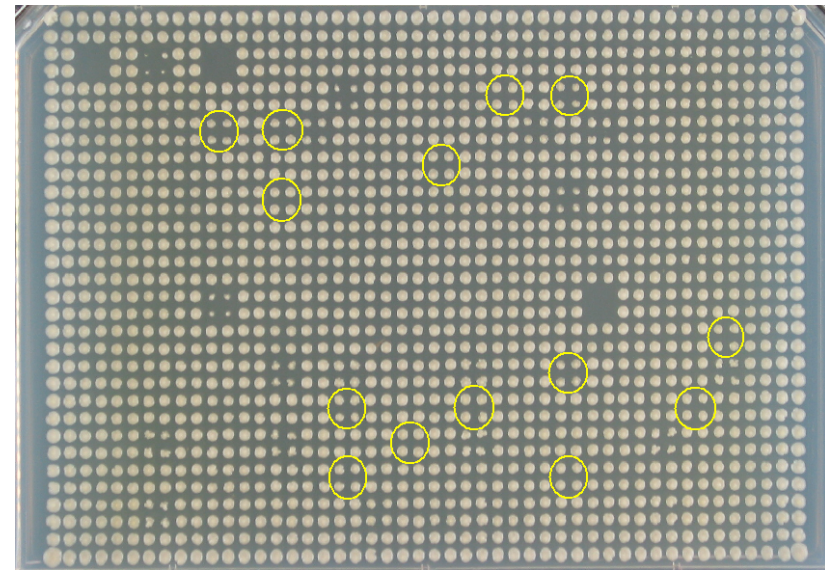
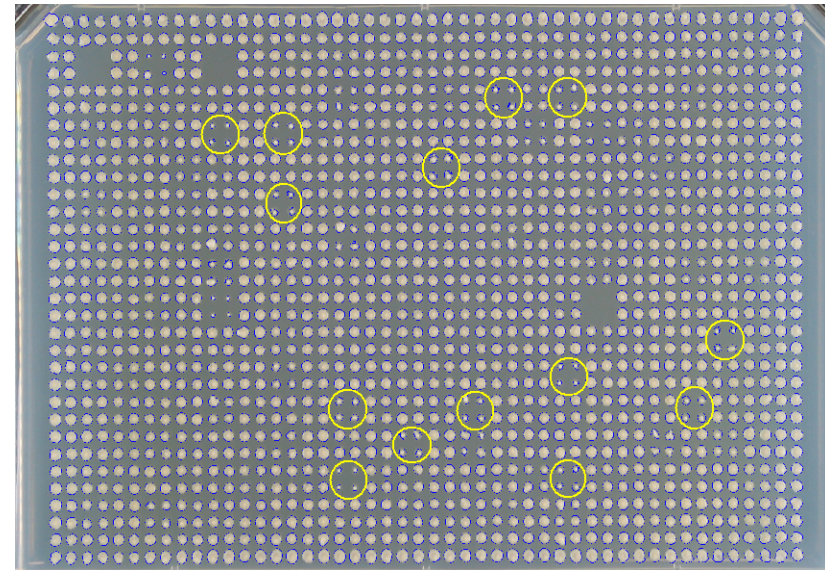
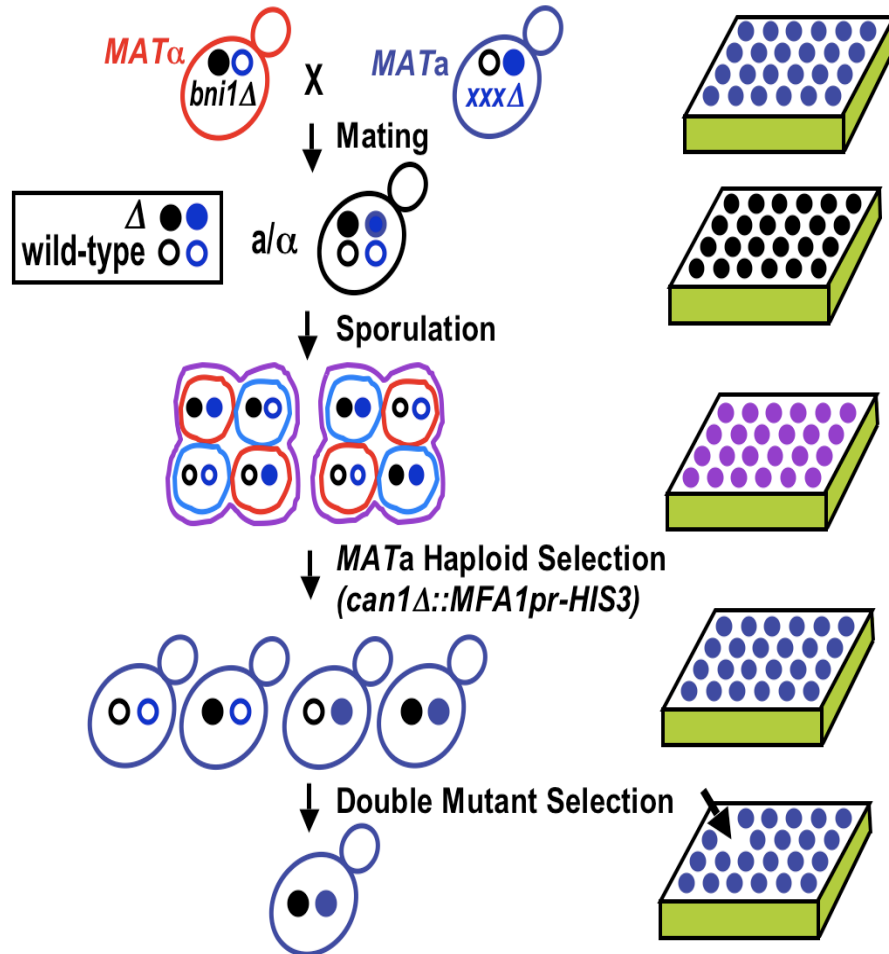


Synthetic Genetic Array (SGA)



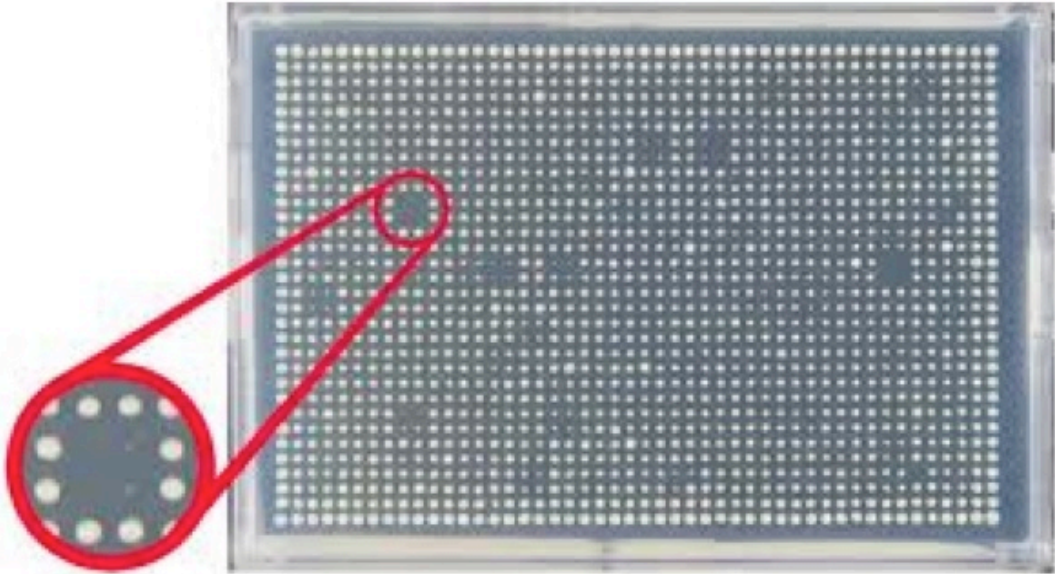
Synthetic Lethal Scoring Program

Query: *VMA6Δ*

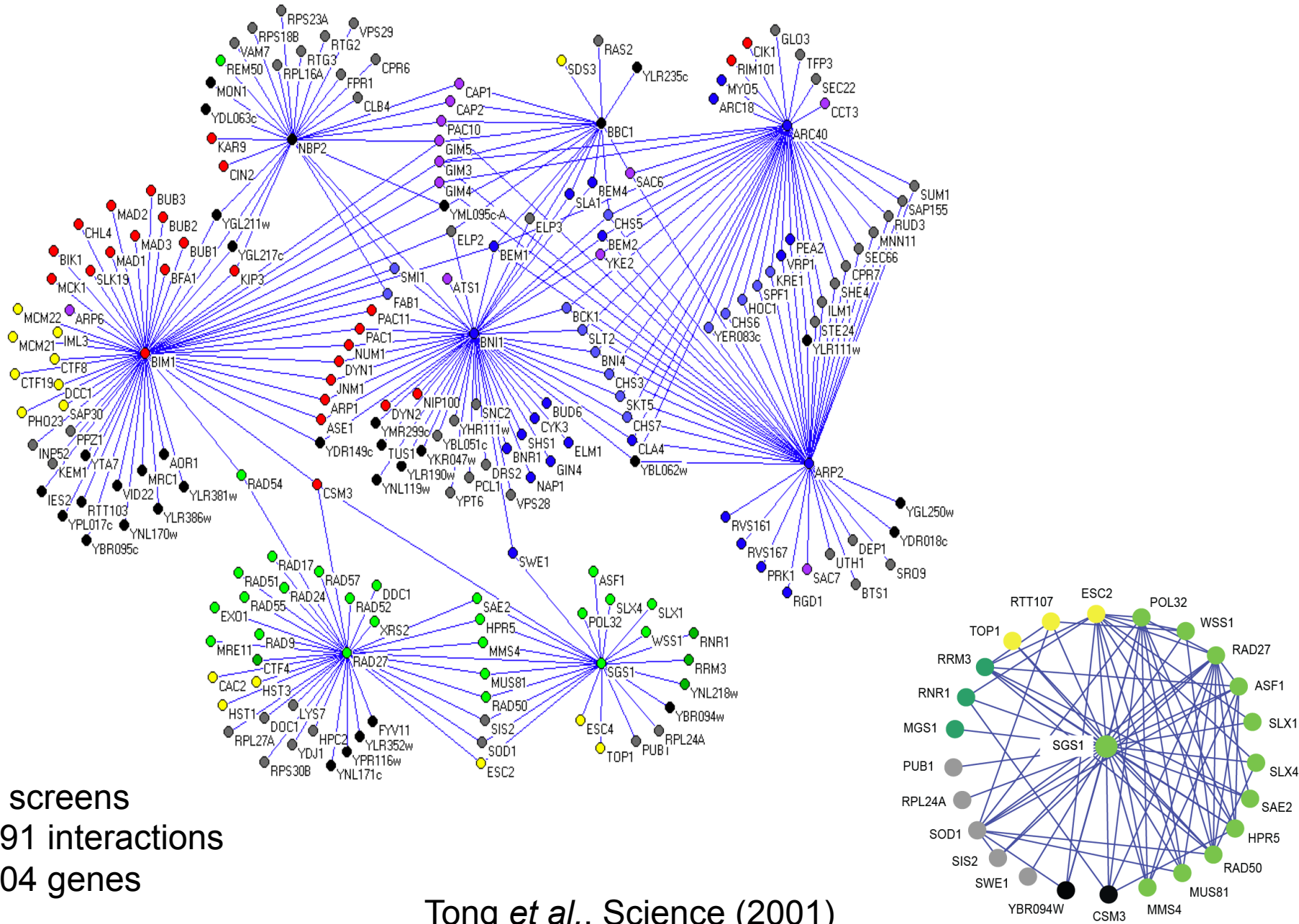


Wildtype control

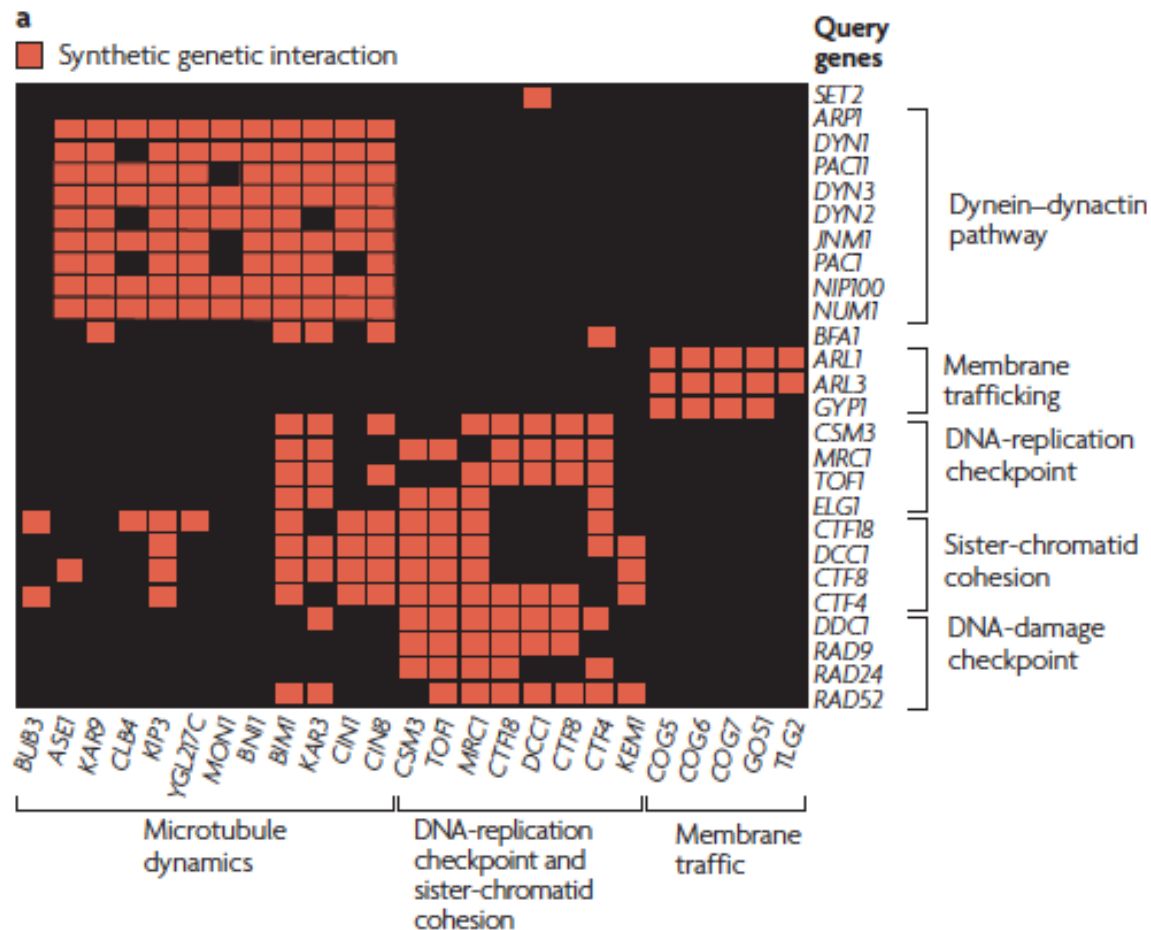
A blow-up, showing the 4-fold replica for each double mutant



First genetic interaction (synthetic lethal) network



Another view of the data: hierarchical clustering



~2000 Quantitative SGA Screens

5.4 million gene-gene pairs, ~30% of Total Network
170,000 Interactions, ~2/3 Negative 1/3 Positive

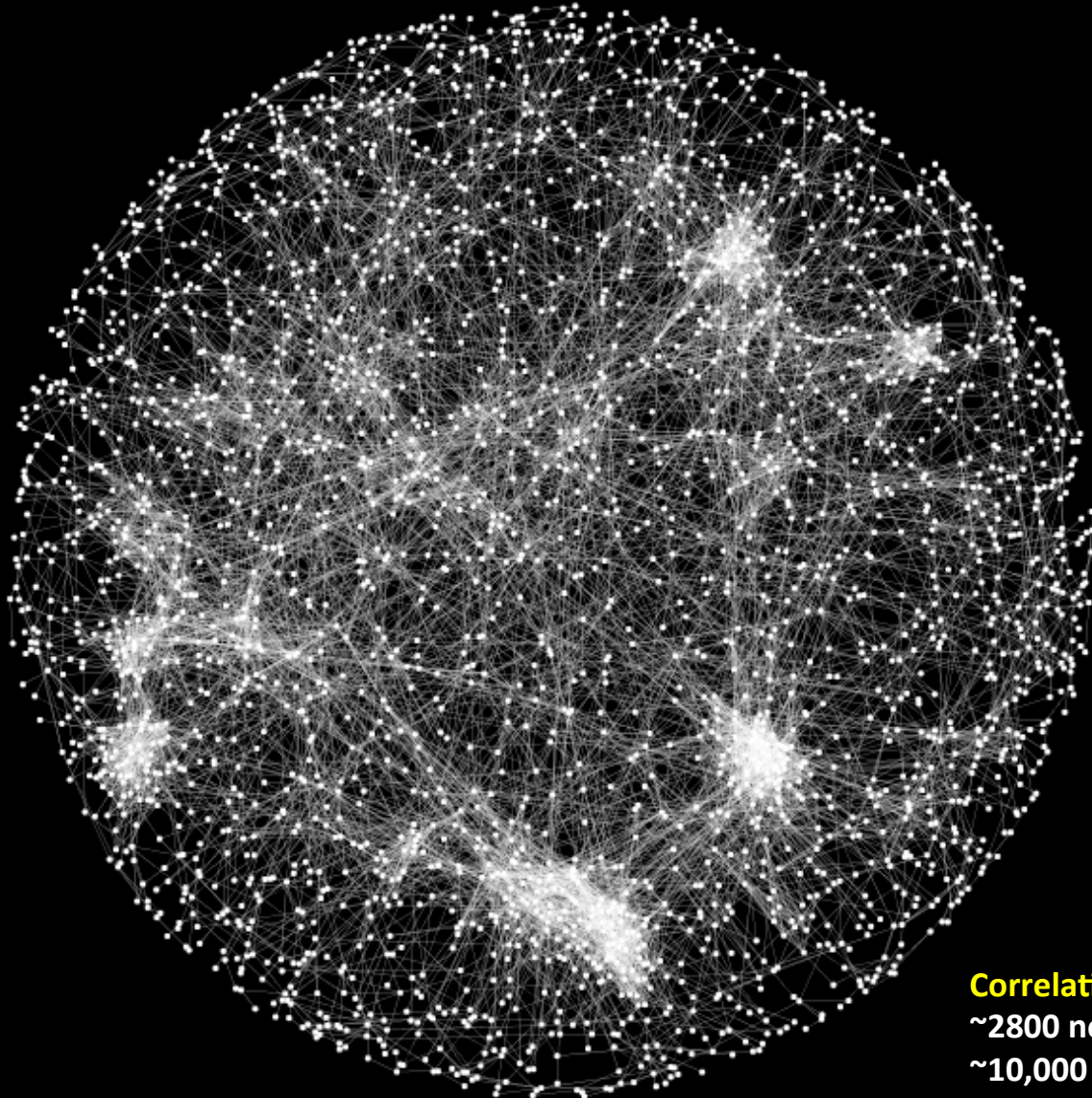
The Genetic Landscape of a Cell

Michael Costanzo,^{1,2*} Anastasia Baryshnikova,^{1,2*} Jeremy Bellay,³ Yungil Kim,³ Eric D. Spear,⁴ Carolyn S. Sevier,⁴ Huiming Ding,^{1,2} Judice L.Y. Koh,^{1,2} Kiana Toufighi,^{1,2} Sara Mostafavi,^{1,5} Jeany Prinz,^{1,2} Robert P. St. Onge,⁶ Benjamin VanderSluis,³ Taras Makhnevych,⁷ Franco J. Vizeacoumar,^{1,2} Solmaz Alizadeh,^{1,2} Sondra Bahr,^{1,2} Renee L. Brost,^{1,2} Yiqun Chen,^{1,2} Murat Cokol,⁸ Raamesh Deshpande,³ Zhijian Li,^{1,2} Zhen-Yuan Lin,⁹ Wendy Liang,^{1,2} Michaela Marback,^{1,2} Jadine Paw,^{1,2} Bryan-Joseph San Luis,^{1,2} Ermira Shuteriqi,^{1,2} Amy Hin Yan Tong,^{1,2} Nydia van Dyk,^{1,2} Iain M. Wallace,^{1,2,10} Joseph A. Whitney,^{1,5} Matthew T. Weirauch,¹¹ Guoqing Zhong,^{1,2} Hongwei Zhu,^{1,2} Walid A. Houry,⁷ Michael Brudno,^{1,5} Sasan Ragibizadeh,¹² Balázs Papp,¹³ Csaba Pál,¹³ Frederick P. Roth,⁸ Guri Giaever,^{2,10} Corey Nislow,^{1,2} Olga G. Troyanskaya,¹⁴ Howard Bussey,¹⁵ Gary D. Bader,^{1,2} Anne-Claude Gingras,⁹ Quaid D. Morris,^{1,2,5} Philip M. Kim,^{1,2} Chris A. Kaiser,⁴ Chad L. Myers,^{3†} Brenda J. Andrews,^{1,2†} Charles Boone^{1,2†}

A genome-scale genetic interaction map was constructed by examining 5.4 million gene-gene pairs for synthetic genetic interactions, generating quantitative genetic interaction profiles for ~75% of all genes in the budding yeast, *Saccharomyces cerevisiae*. A network based on genetic interaction profiles reveals a functional map of the cell in which genes of similar biological processes cluster together in coherent subsets, and highly correlated profiles delineate specific pathways to define gene function. The global network identifies functional cross-connections between all bioprocesses, mapping a cellular wiring diagram of pleiotropy. Genetic interaction degree correlated with a number of different gene attributes, which may be informative about genetic network hubs in other organisms. We also demonstrate that extensive and unbiased mapping of the genetic landscape provides a key for interpretation of chemical-genetic interactions and drug target identification.

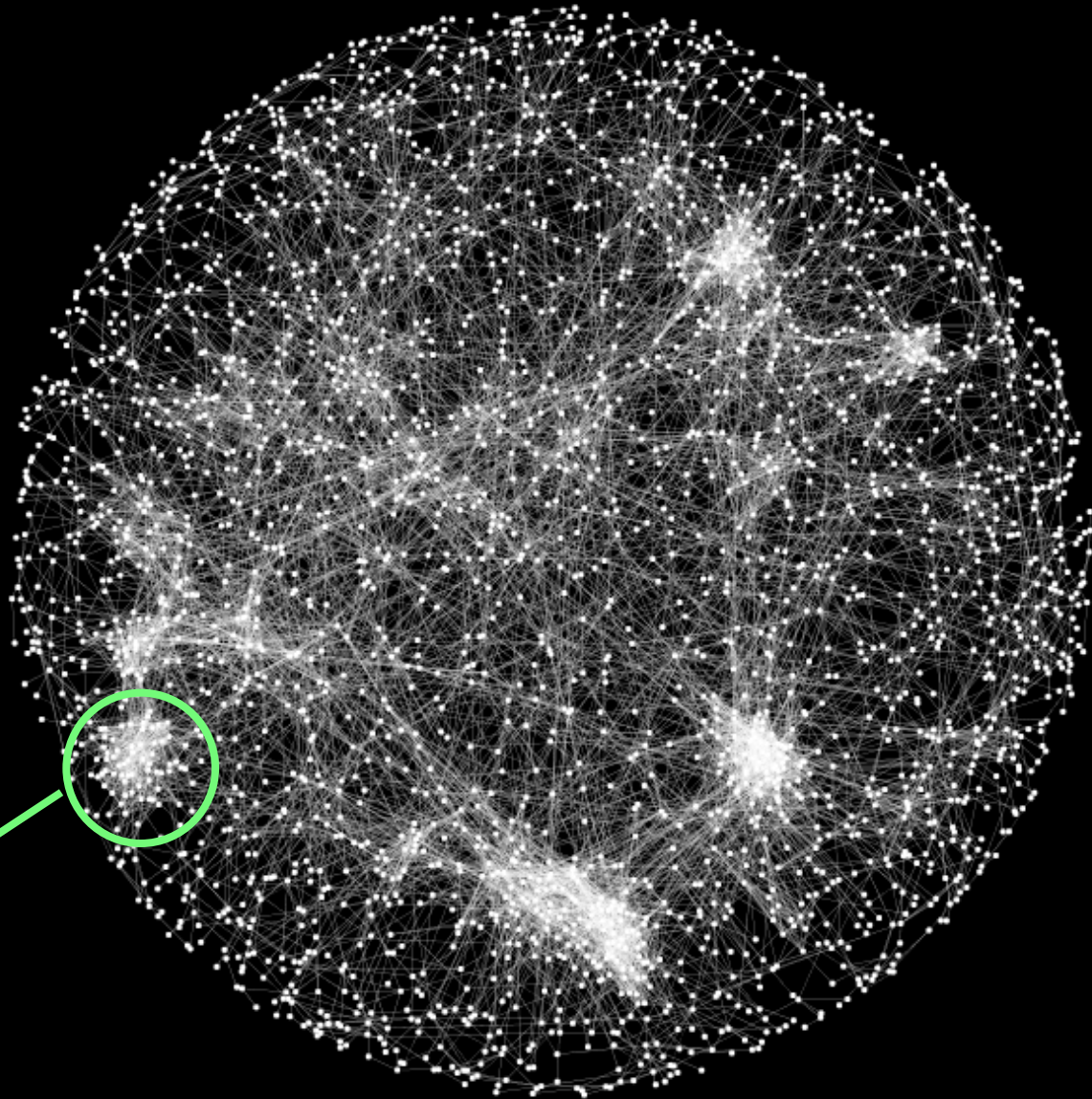


Yeast Genetic Interaction Network Global Level



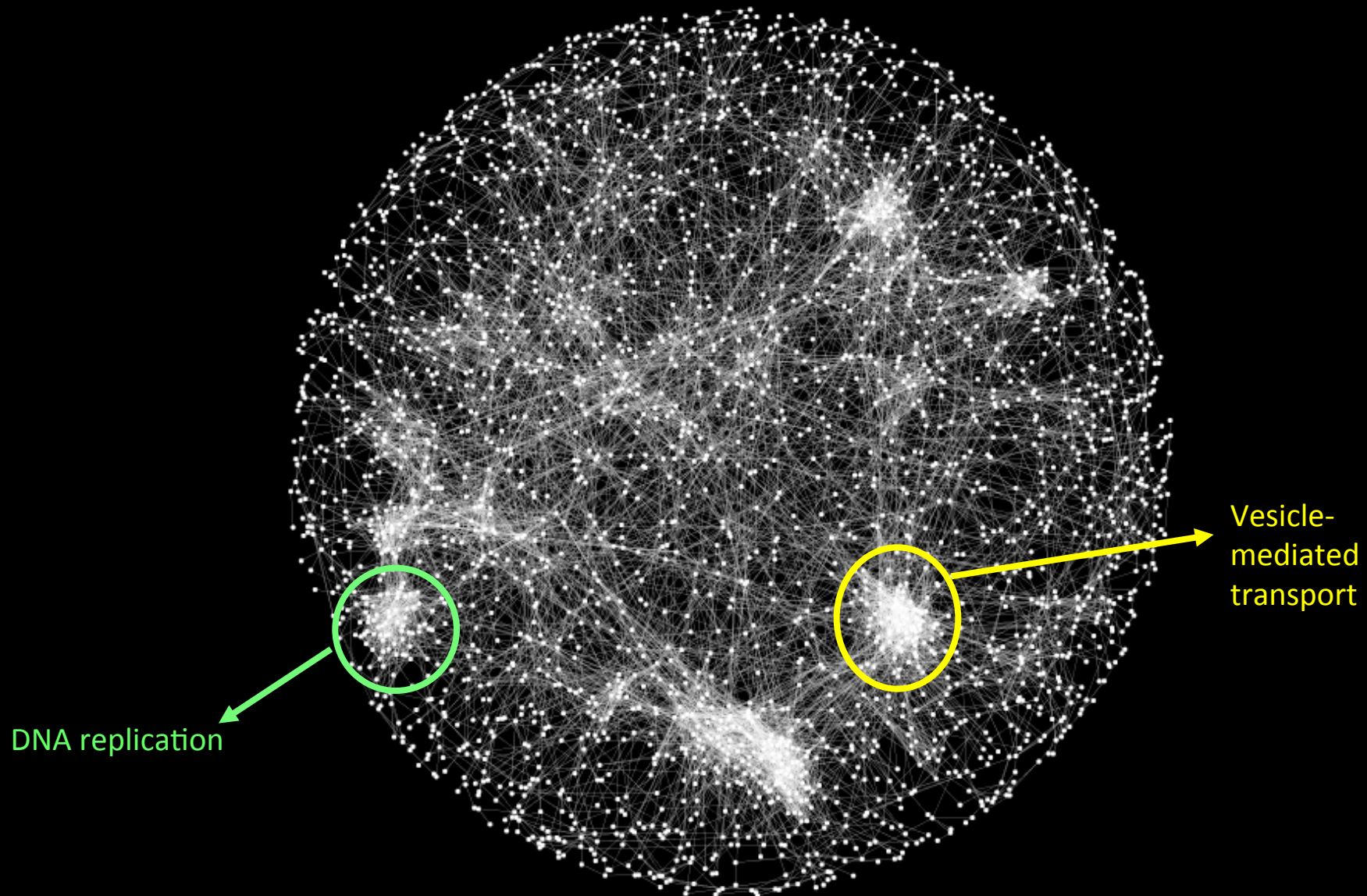
Correlation-based network
~2800 nodes
~10,000 correlation edges

Yeast Genetic Interaction Network Global Level

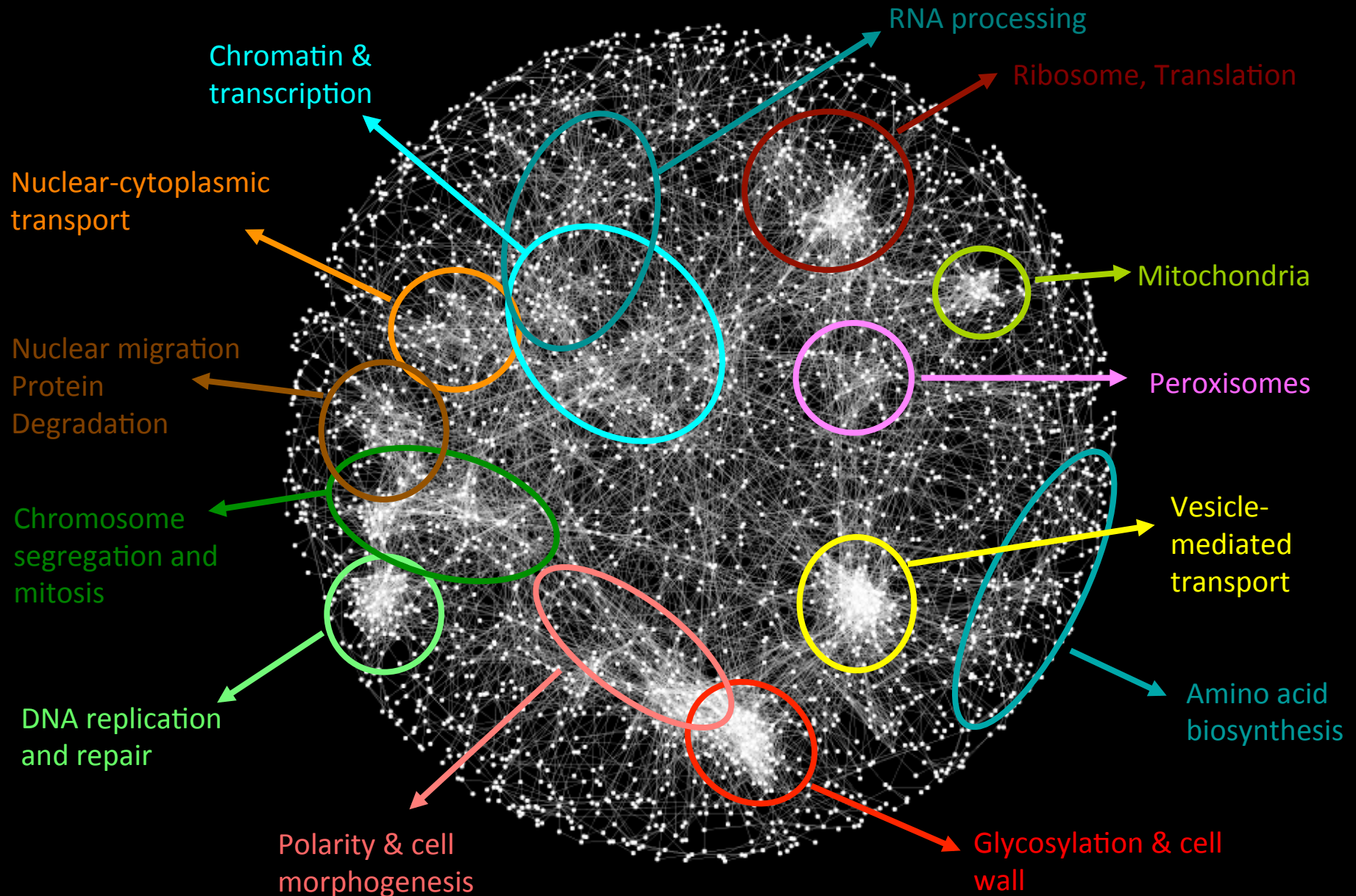


DNA replication

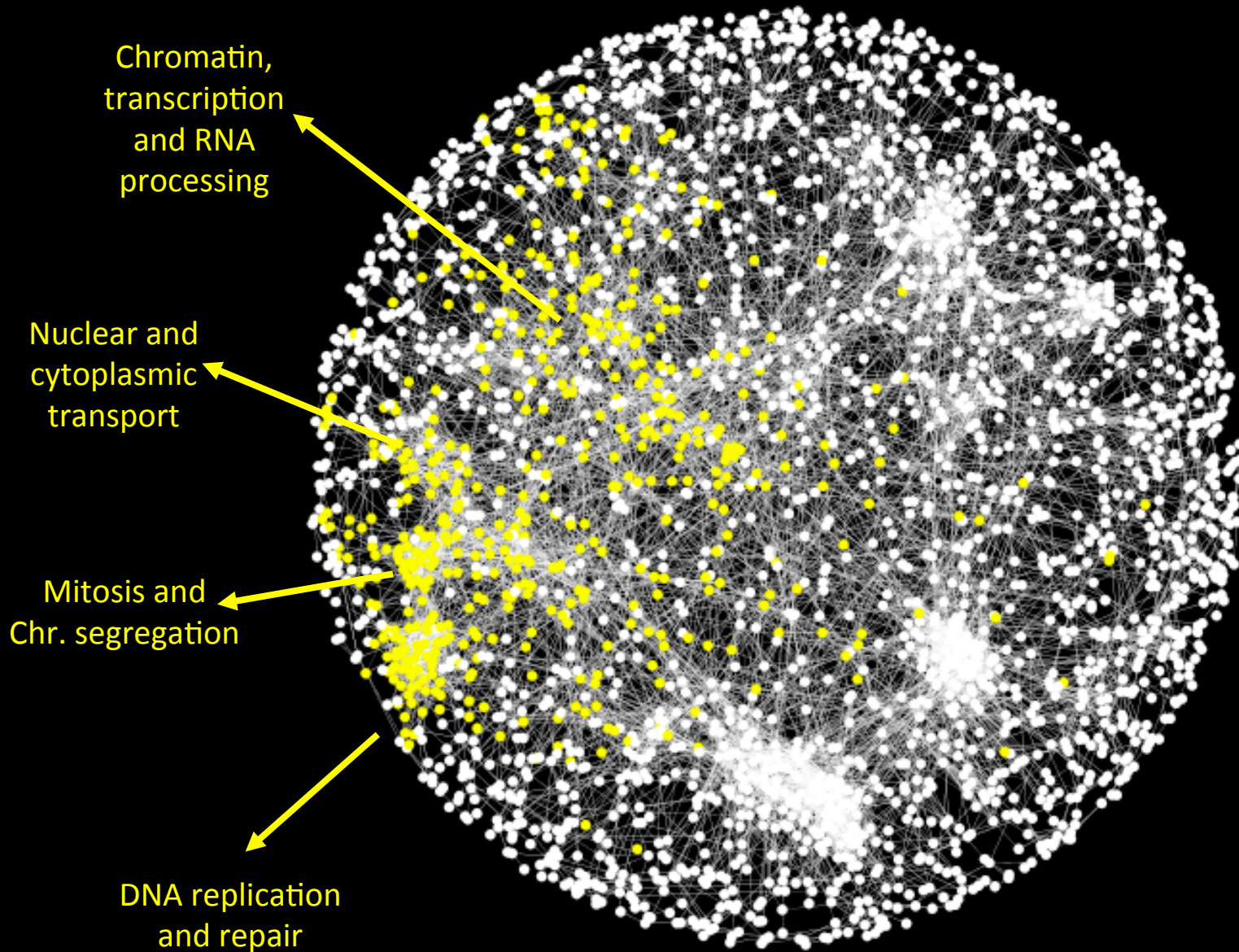
Yeast Genetic Interaction Network Global Level



Yeast Genetic Interaction Network Global Level

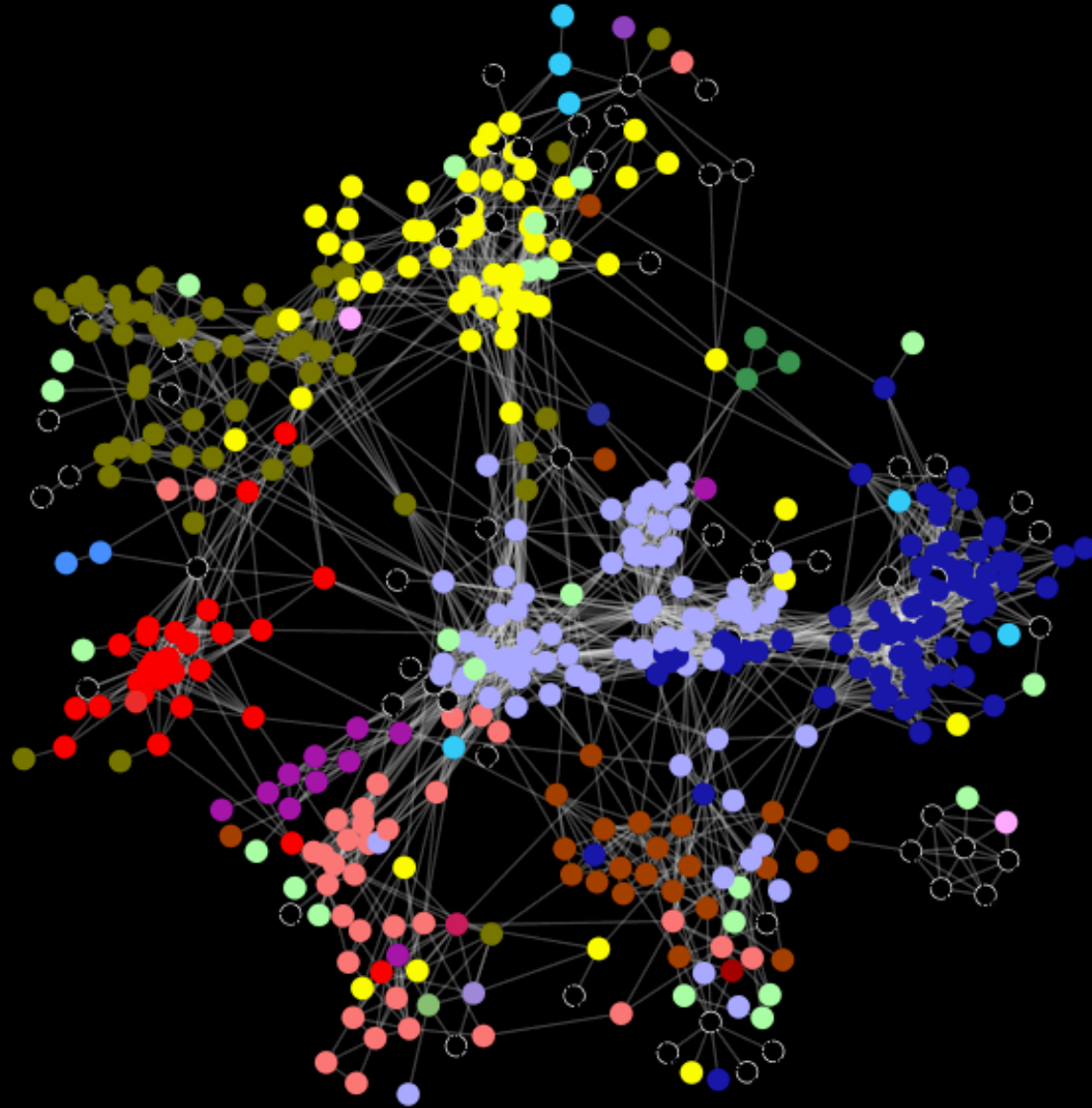
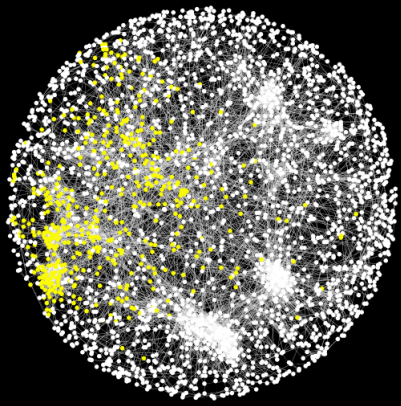


Yeast Genetic Interaction Network Global Level



Yeast Genetic Interaction Network

Process Level





DRYGIN



Data Repository of Yeast Genetic Interactions

Quick gene search:
enter gene names or orfs

[Problem with searching?](#)

[Home](#) | [Search genes](#) | [Sub-networks](#) | [Search complexes/Pathways](#) | [Overlap with BIOGRID](#) | [Download](#) | [About DRYGIN](#)

DRYGIN is a database of quantitative genetic interactions of *S. Cerevisiae* derived from the SGA double-mutant arrays conducted in [Boone lab](#) at [Terrence Donnelly Centre for Cellular and Biochemical Research, University of Toronto](#).

Current version of DRYGIN is 1.0 ([DRYGIN documentation](#)).

Latest Database statistics

- Total SGA genetic interactions in DRYGIN: 5,482,948
- Total genes screened: 1,711 query (1,672 ORFs) X 3,885 array (3,885 ORFs)
- **Updates:**
 - 2010-02-10: The database was updated with the latest values from Costanzo et al., 2010.
 - 2010-11-18: The *clc1Δ* screen was removed because an allele discrepancy was noticed.

For description of the database content, please refer to [Judice L.Y. Koh, Huiming Ding, Michael Costanzo, Anastasia Baryshnikova, Kiana Toufighi, Gary D. Bader, Chad L. Myers, Brenda J. Andrews, and Charles Boone](#). DRYGIN: a database of quantitative genetic interaction networks in yeast. *Nucleic Acids Research*, 2010, Vol. 38, Database issue D502-D507.



[Search genes](#) - Query SGA genetic interactions using gene names or orfs. Download interactions or visualize network or clustergram, with genes color-coded according to functional annotations.



[Sub-networks](#) - Search and visualize sub-networks of genetic interactions between a group of genes.



[Search complexes/pathways](#) - Search and visualize SGA genetic interactions in known protein complexes and pathways.



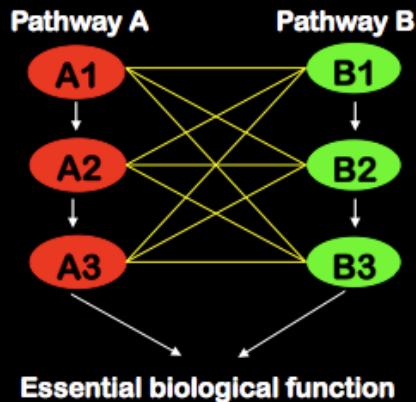
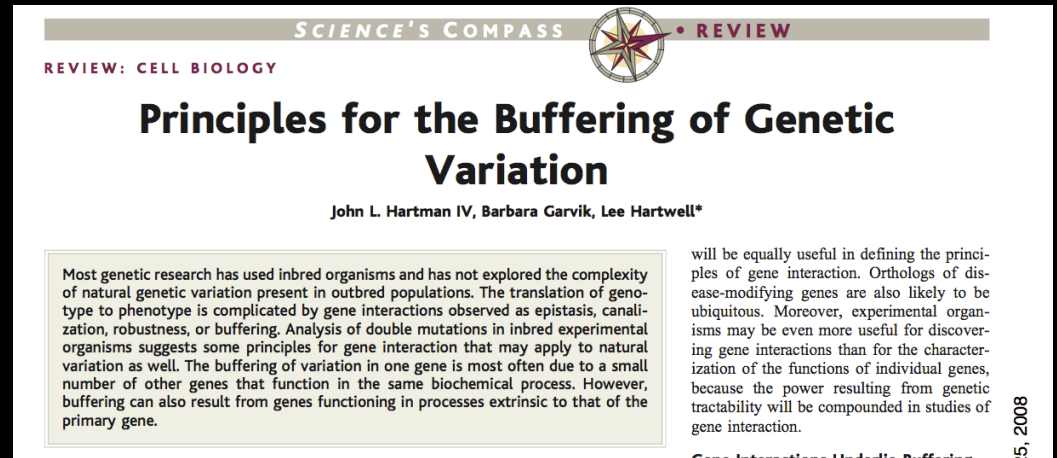
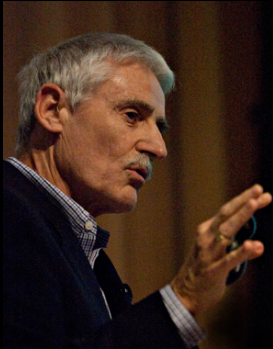
[Overlap with BIOGRID](#) - Search for SGA genetic interactions that overlap with BIOGRID genetic and physical interactions.



[Download](#) - Download latest SGA genetic interactions in Cytoscape SIF, tab-delimited, or Java Treeview CDT formats.

The Hartwell Idea

Synthetic Lethal Networks May Guide Our Understanding of Genotype to Phenotype



Hartman, Garvik, and Hartwell,
Science, 9 February (2001), Pg. 1001

“analysis of double mutations in inbred experimental organisms suggest principles that may apply to natural variation in outbred populations”

The Fink Idea

Σ1278b

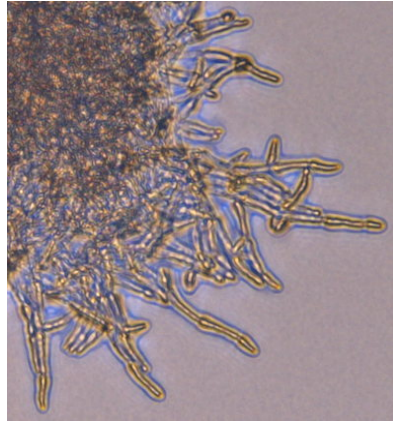


S288c

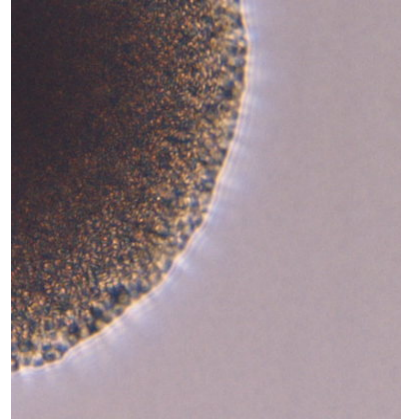


Genotype/Phenotype of *S. cerevisiae* Strains

Σ1278b



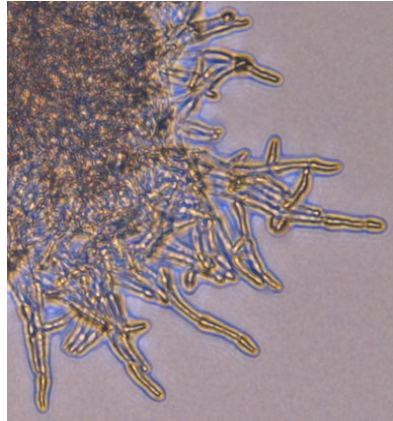
S288c



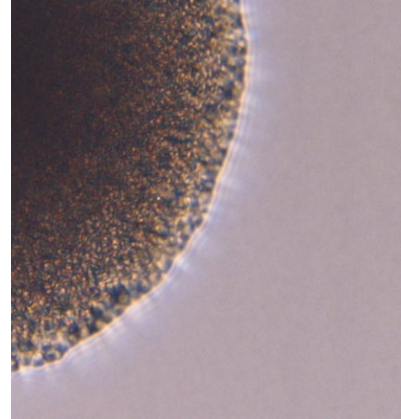
- **Strains Mate, Meiotic Progeny are Viable**

Genotype/Phenotype of *S. cerevisiae* Strains

Σ 1278b



S288c



- **Strains Mate, Meiotic Progeny are Viable**
 - **G. Fink, R Dowell, and D. Gifford (MIT) sequenced Σ 1278b.**
- ~0.2% natural variation between strains**

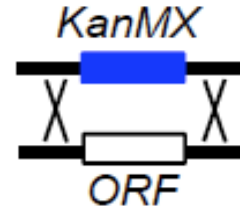


Σ1278b Deletion Mutant Collection

Exploring Conditional or Background-specific Phenotypes

A. Σ1278b Deletion Mutant Collection

1. Transform
Barcoded *KanMX*
Deletion Alleles



2. 5127 Heterozygous
a/α Deletion Mutants



3. Sporulation



4. Test essentiality
by random spore
germination of haploid
MATa meiotic progeny
(*STE2pr-sphis5* selection)



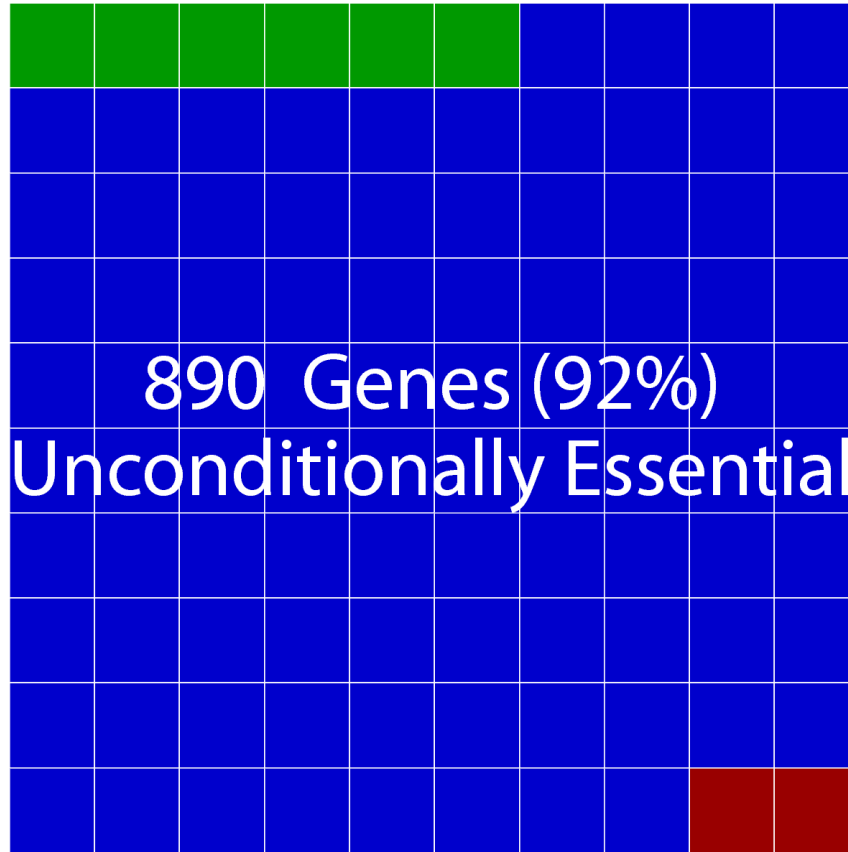
Owen Ryan

Deletion Mutant Analysis in $\Sigma 1278b$ should reveal two kinds of essential genes:

- 1. Unconditional Essentials, those shared by S288c**
- 2. Conditional Essentials, those specific to either $\Sigma 1278b$ or S288c**

Conditional Loci may identify background-specific polymorphisms that cause synthetic lethality.

55 Genes (6%)
Σ1278b Conditionally Essential

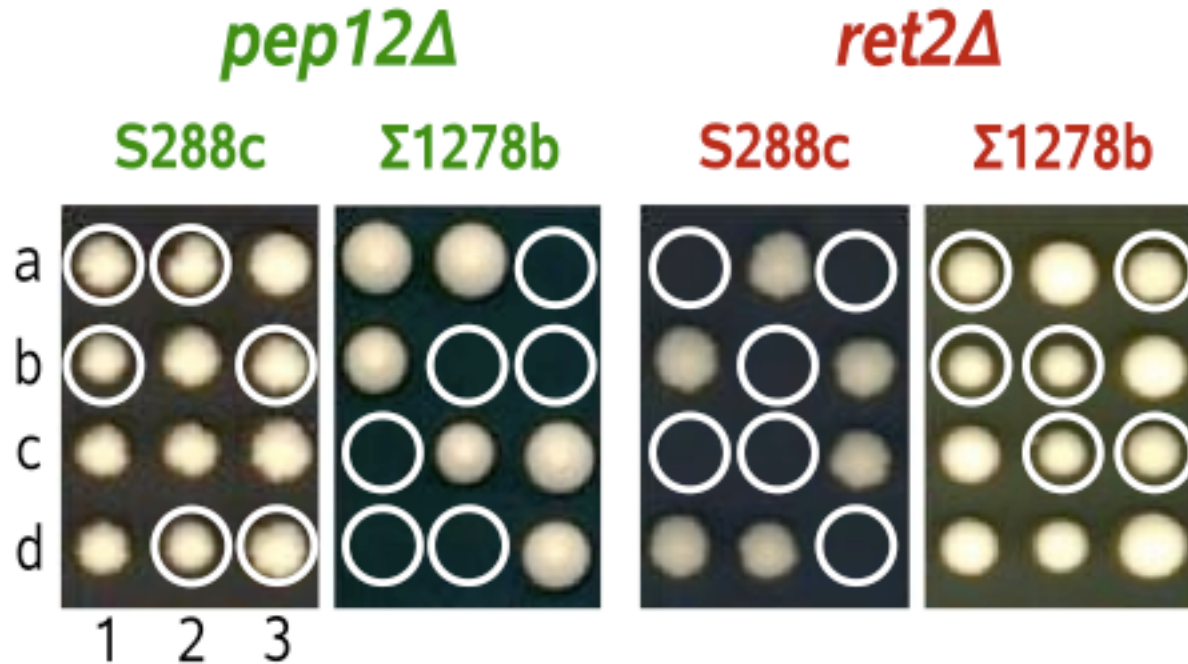


890 Genes (92%)
Unconditionally Essential

20 Genes (2%)
S288c Conditionally Essential

Conditional Essential Genes:

Genes Required for Life in One Individual but not the Other

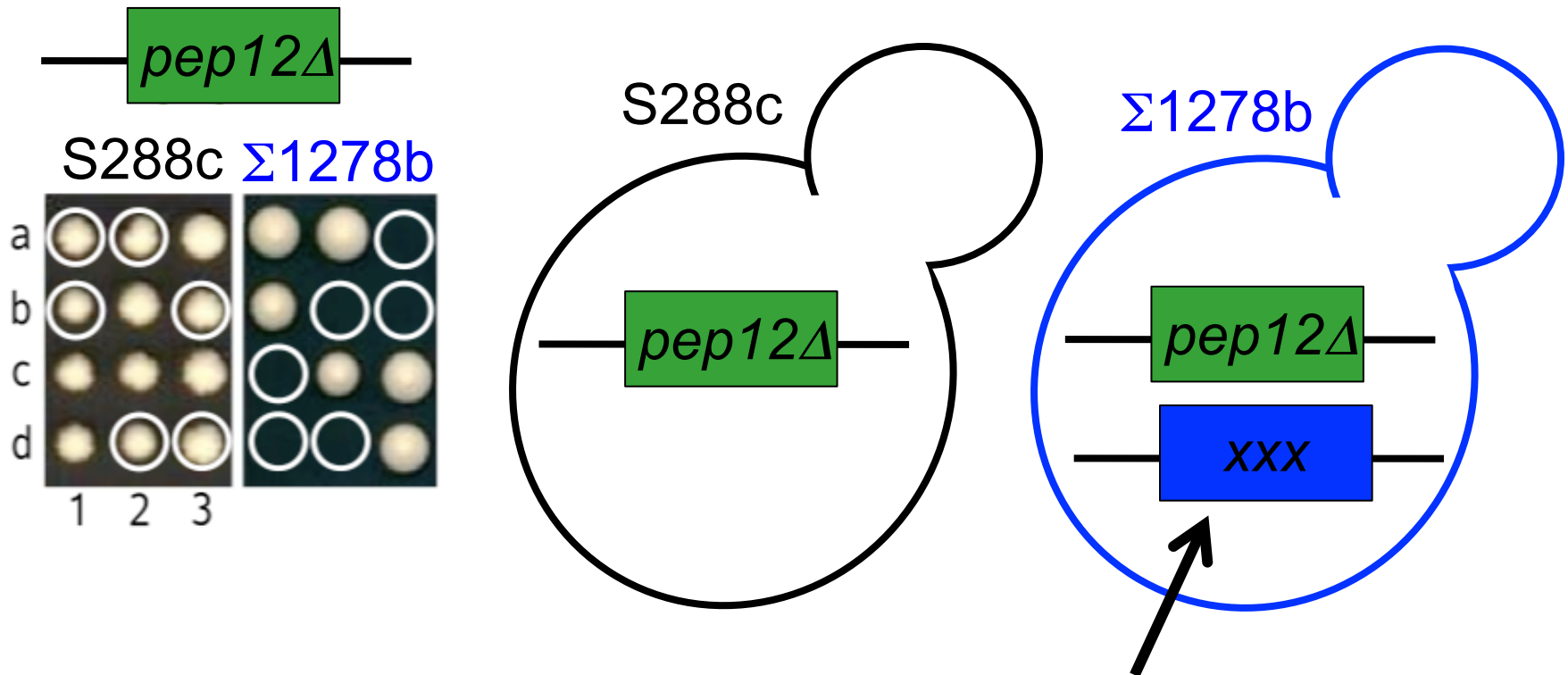


Tetrad dissection: 4 spores (a,b, c, and d) from a single meiosis are separated and allowed to grow into colonies. In this experiment, 2 different diploid genotypes were put through meiosis, *PEP12/pep12* in one case and *RET2/ret2* in the other.

Circles indicate the segregants carrying the deletion allele

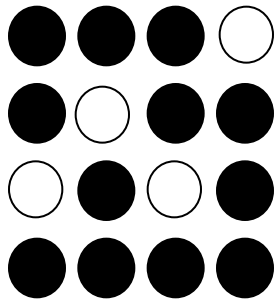
Mechanism of Conditional Lethality

Maybe $\Sigma 1278b$ Carries a Modifier Locus that Leads to Classic Digenic Synthetic Lethality with a *pep12* Δ Mutation

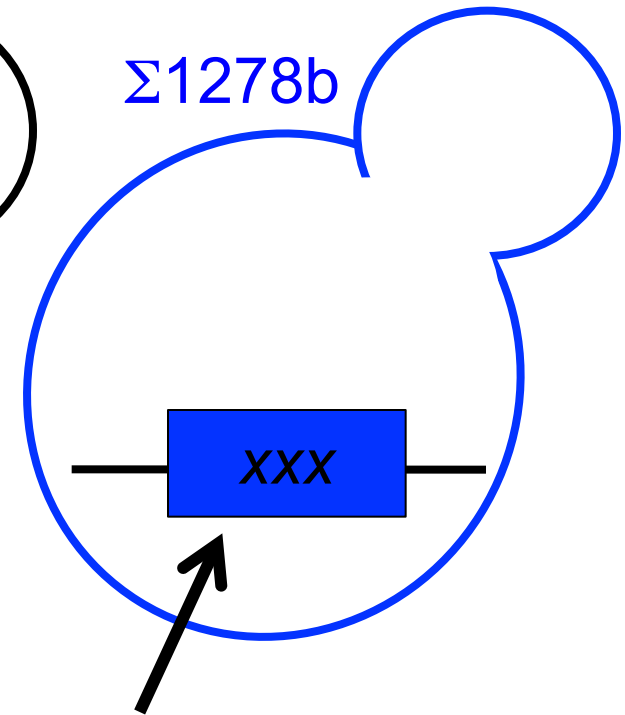
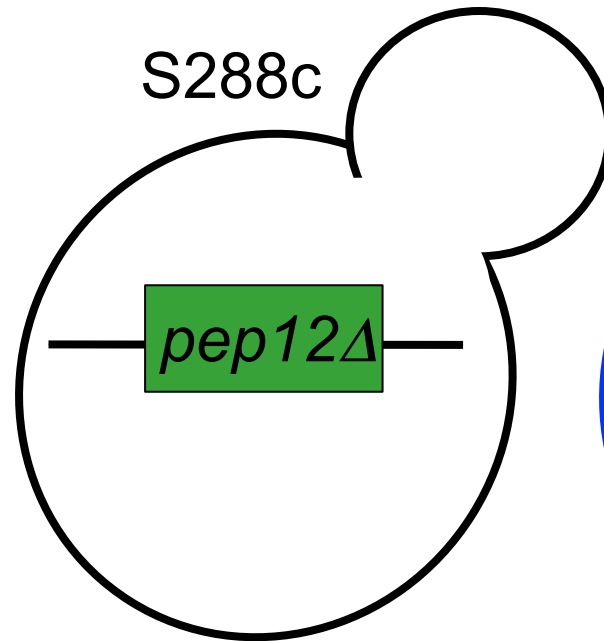


Maybe a Modifier Causes Synthetic Lethality

Testing Number of Modifiers Underlying of Conditional Lethality



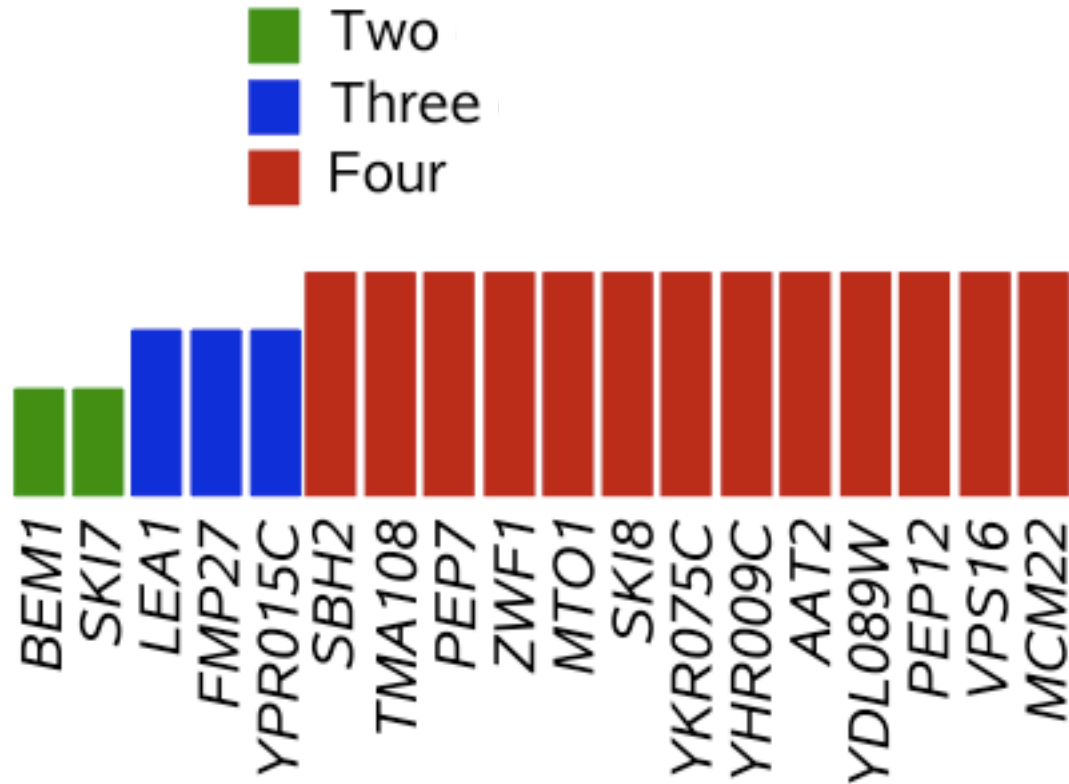
1/4 Spores will die
with 1 synthetic
lethal modifier



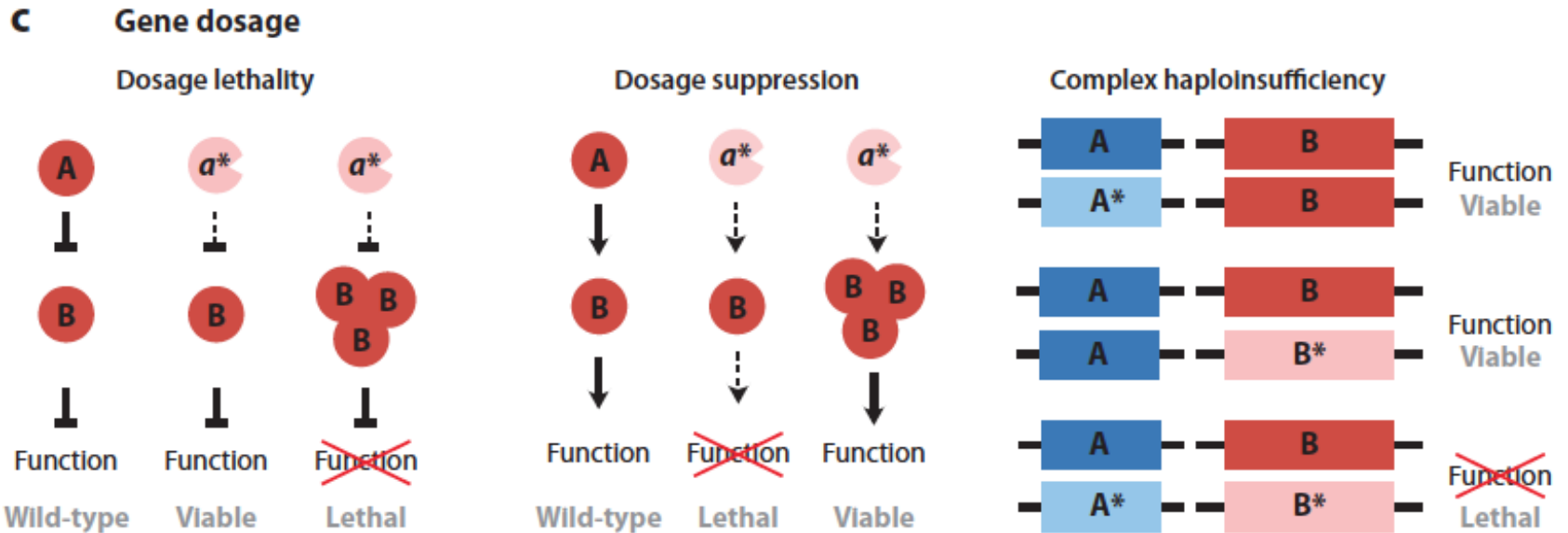
Maybe a Modifier Causes
Synthetic Lethality

Complex Synthetic Lethality Drives Conditional Essentiality

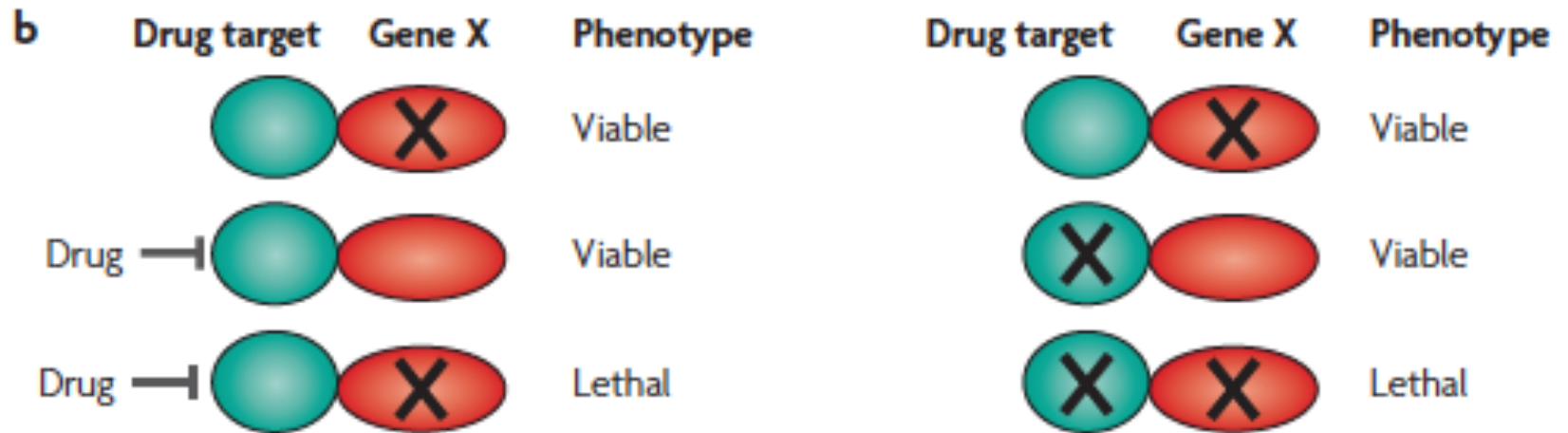
Number of Modifiers



Variations on the synthetic lethal approach



Variations, continued

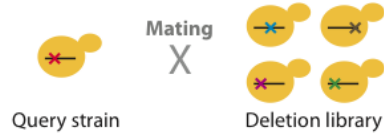


Screen for new drugs or find targets of existing drugs

Systematic genetic screens in yeast, worms and mammalian cells

Step 1: Generate double mutant

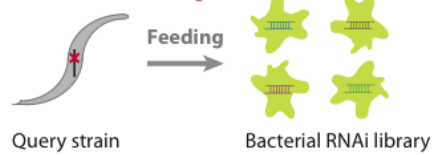
Saccharomyces cerevisiae



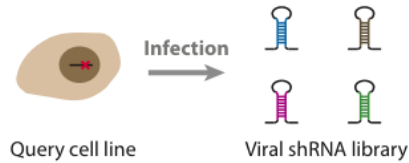
Step 2: Score phenotype and identify interactions



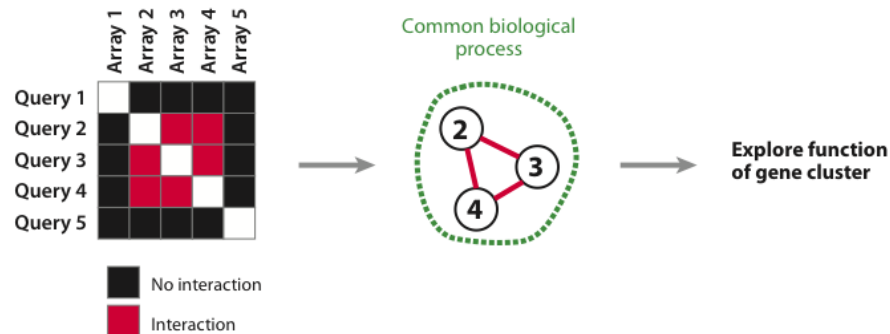
Caenorhabditis elegans



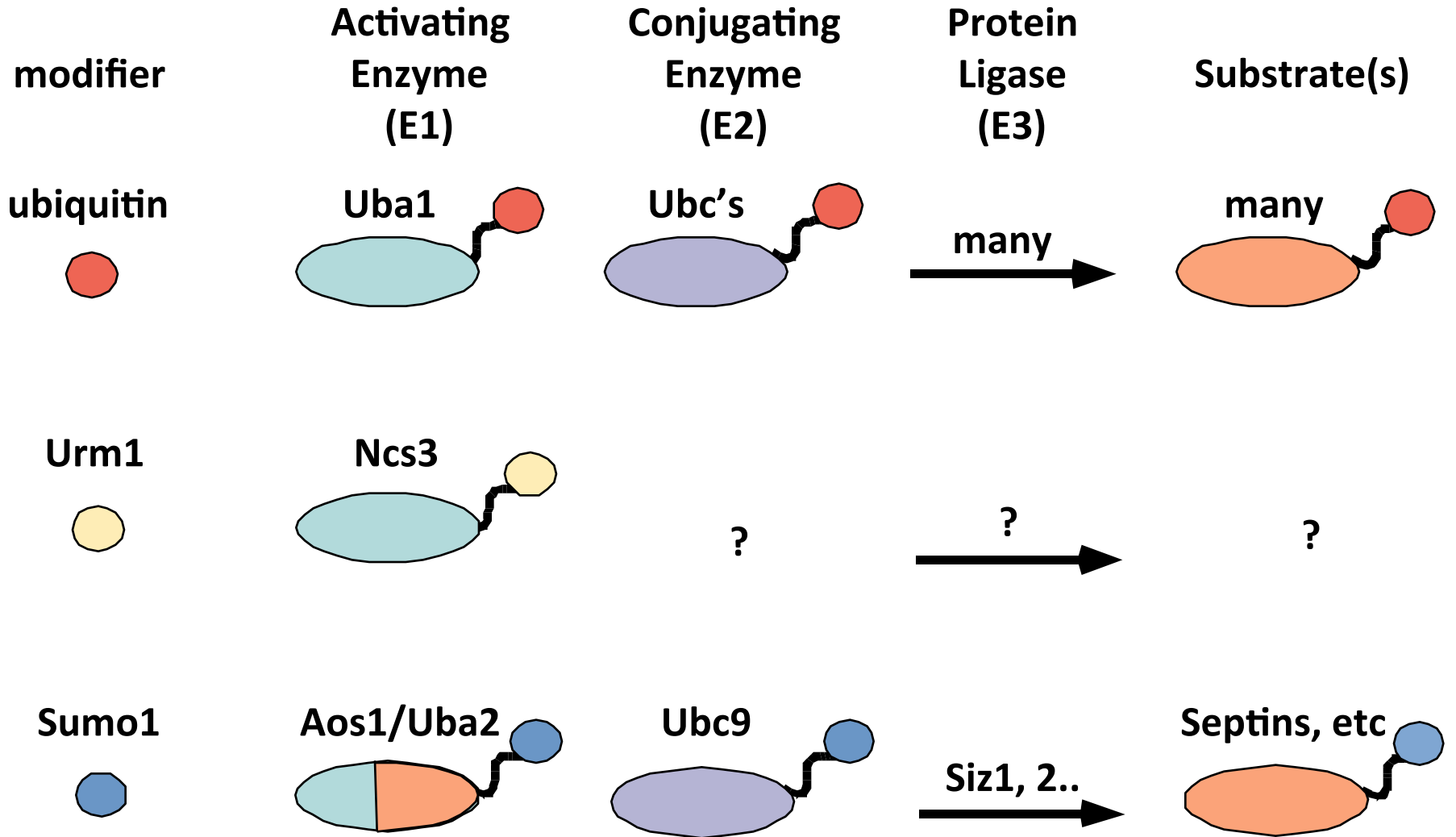
Mammalian cell culture



Step 3: Build genetic interaction networks

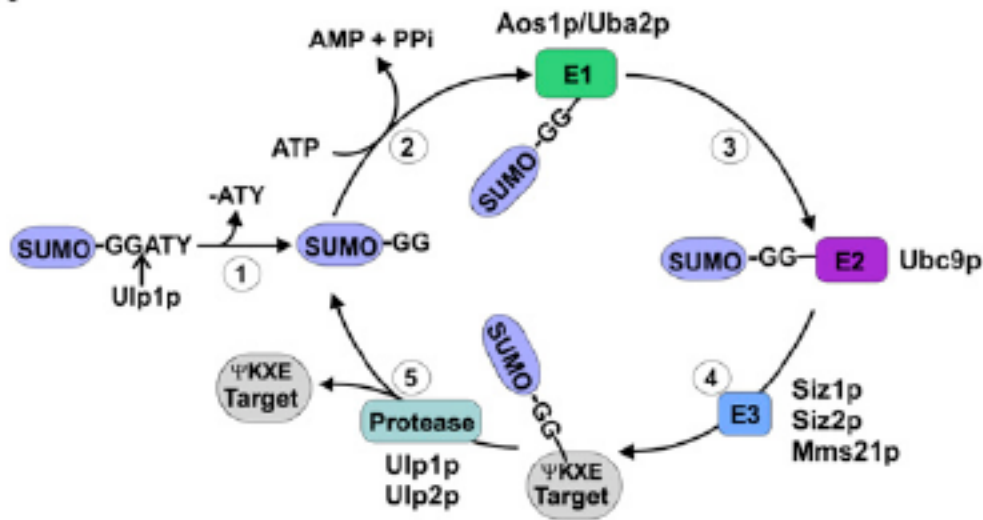


Ubiquitin and Ubiquitin like conjugation pathways

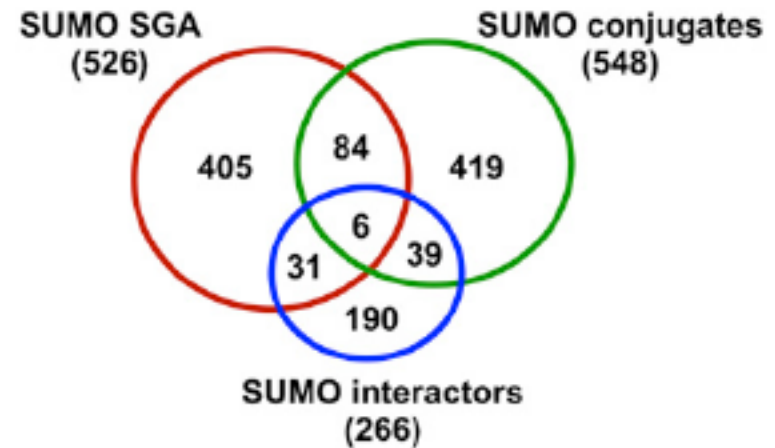


SUMO pathway and “interactors”

A

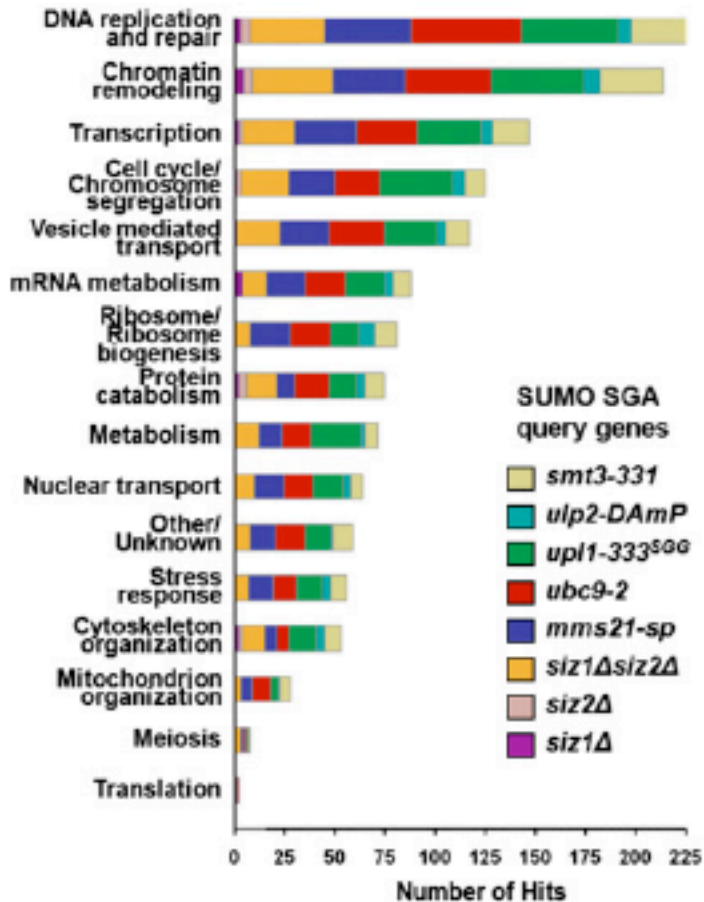


C

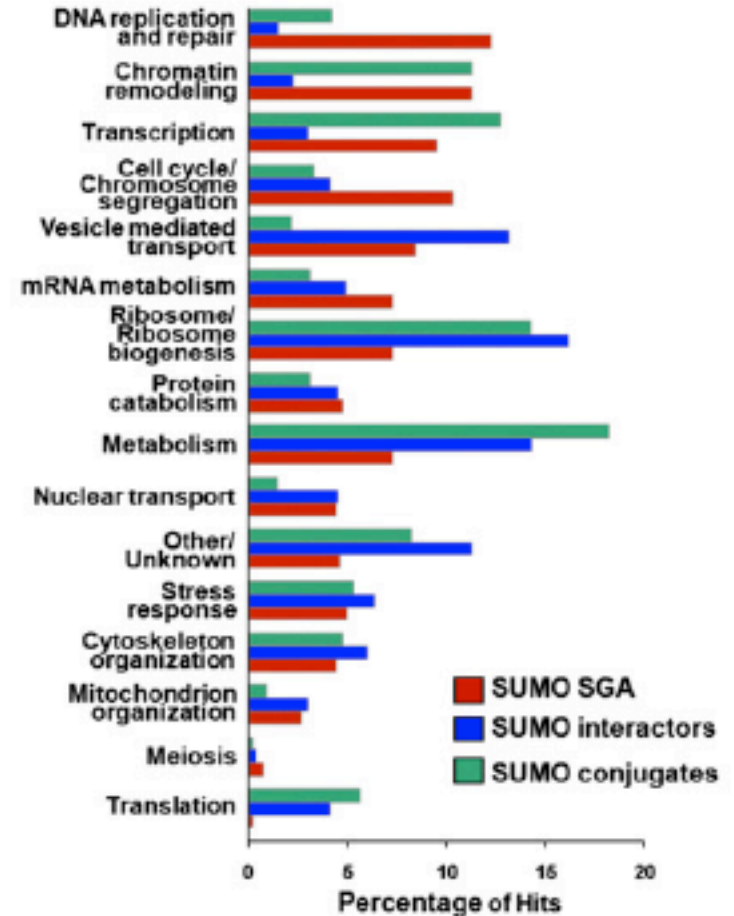


SGA analysis using different SUMO pathway mutants yields similar results

B



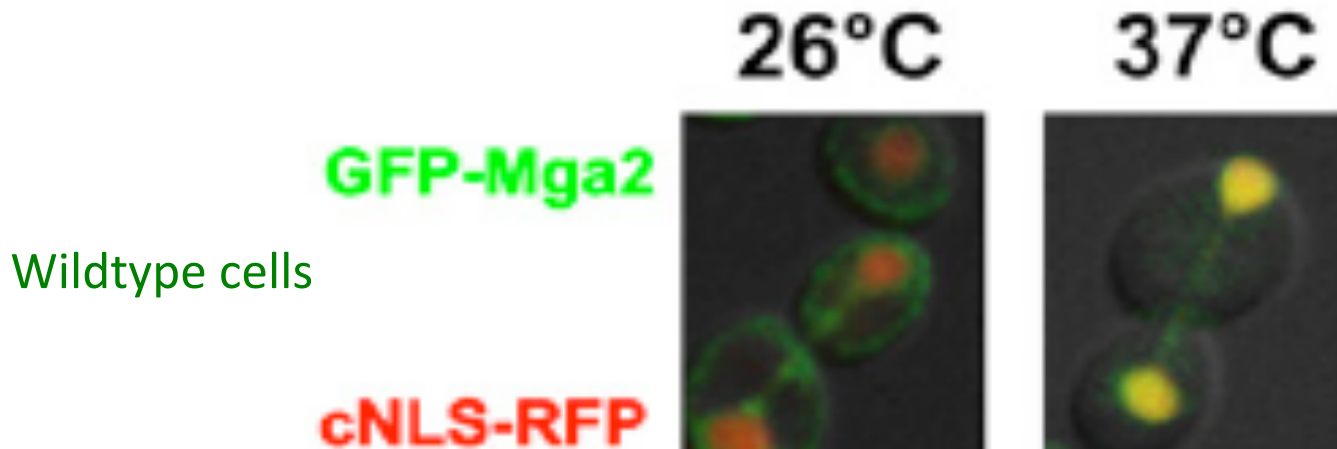
D



But biological processes are represented differently in the 3 analysis methods

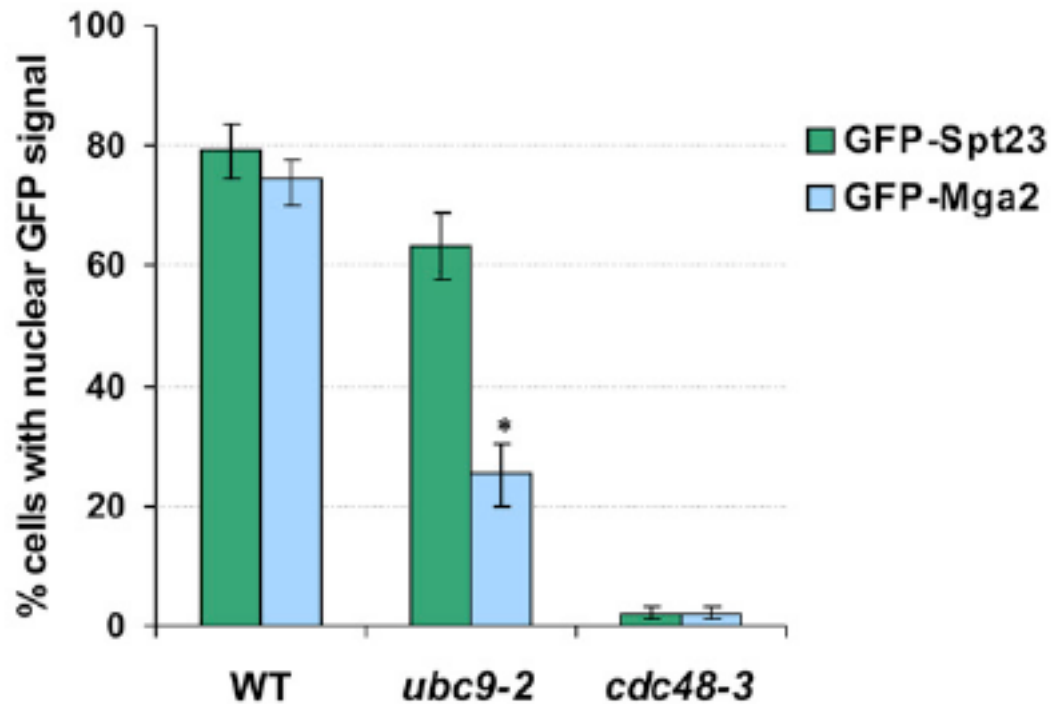
SUMO regulates nuclear localization of a transcription factor

A



C

Quantitation in wildtype and mutants



Haploinsufficiency and complex haploinsufficiency

Haploinsufficiency

In a diploid, “wildtype” allele on one chromosome and loss-of-function allele on the other results in a phenotype

Hence, haploinsufficiency is a kind of dominance. How would you distinguish this sort of dominant allele from a gain-of-function dominant allele?

Complex Haploinsufficiency

In a diploid, loss-of-function alleles at two (or more) different loci, eg $GENEX^+/genex^-$ $GENEY^+/geney^-$, confers a phenotype whereas neither single loss-of-function allele does. The thinking is that complex haploinsufficiency may be akin to the genetics of disease susceptibility in humans

Actin has many roles in the cell

Membrane trafficking

Polarized cell growth

Cytokinesis

Organelle positioning and segregation

Nuclear structure

More

Complex Haploinsufficiency Tests with Actin

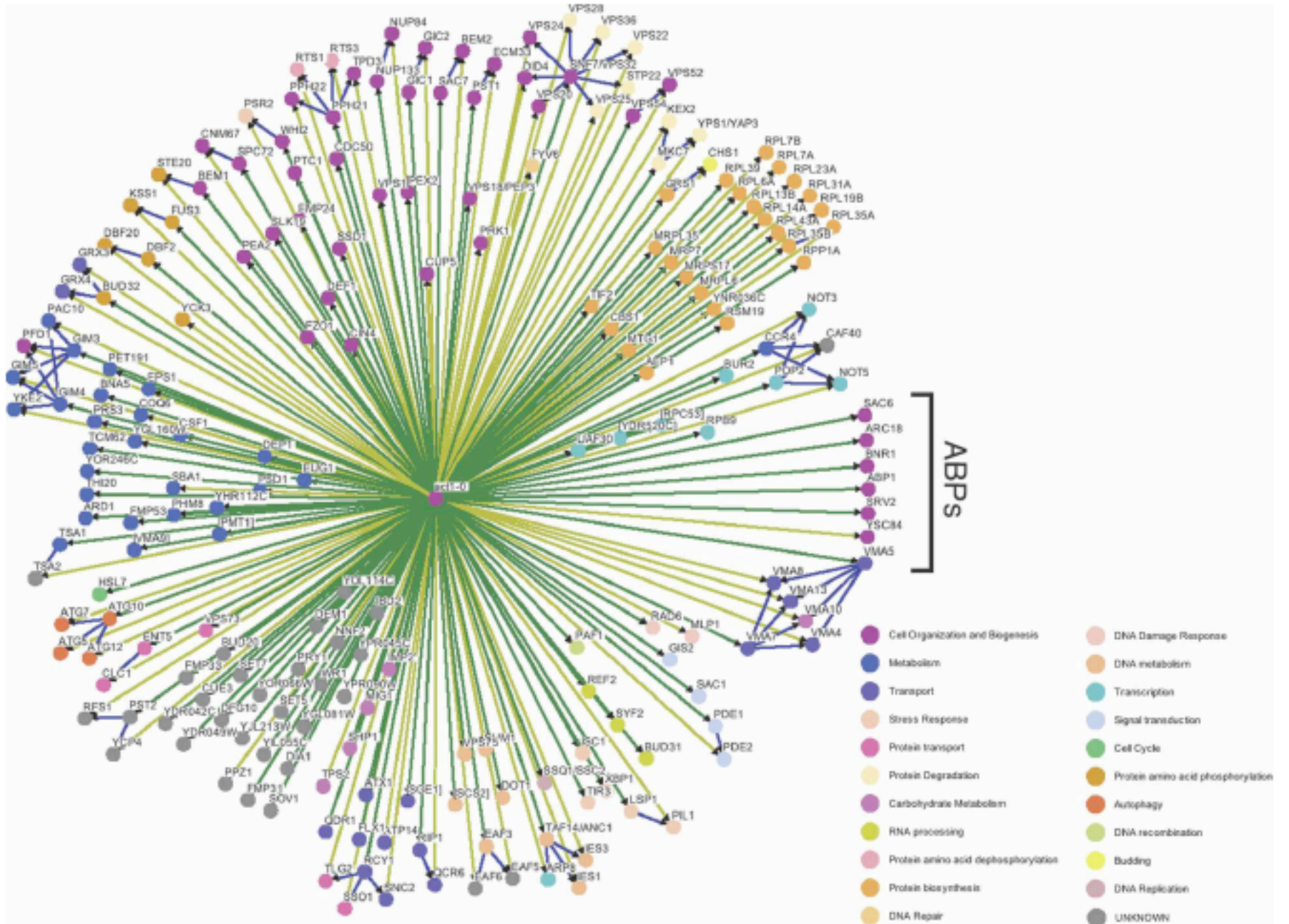
*act1*Δ strain

Carrying plasmid-borne wildtype *ACT1* gene

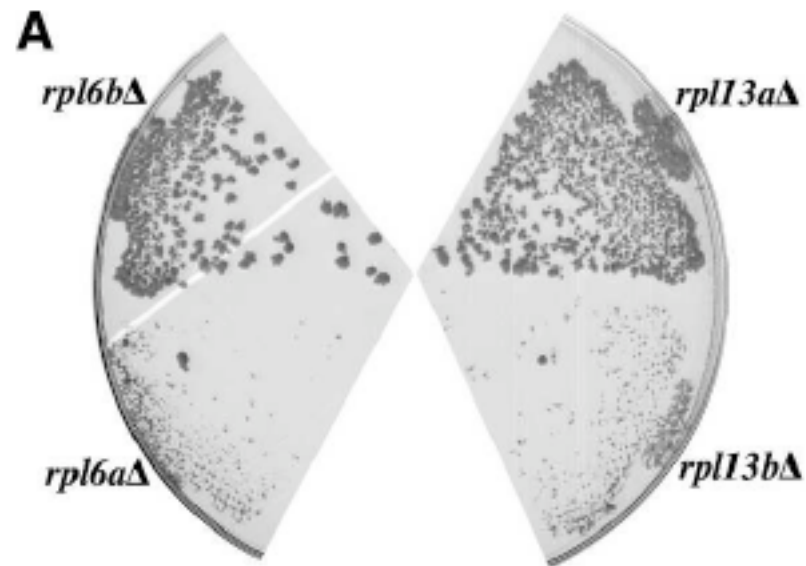
Mate to deletion collection of nonessential genes and select diploids

Determine whether each resulting diploid strain can lose the plasmid.
If not, a CHI between actin and the corresponding gene in the deletion array is suggested

Actin shows complex haploinsufficiency with many nonessential genes



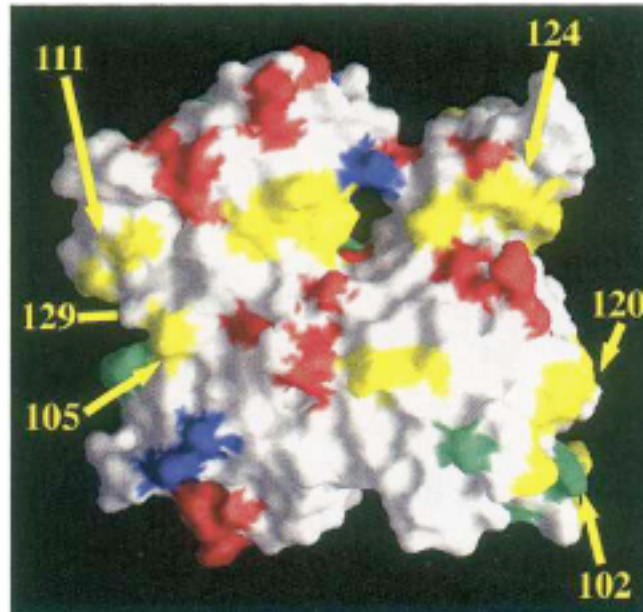
Ribosome large subunit paralogs show different interactions with actin



Genotypes: heterozygous for deletion of *ACT1*
and for each of the indicated genes

Positions of actin substitutions used in haploinsufficiency analysis

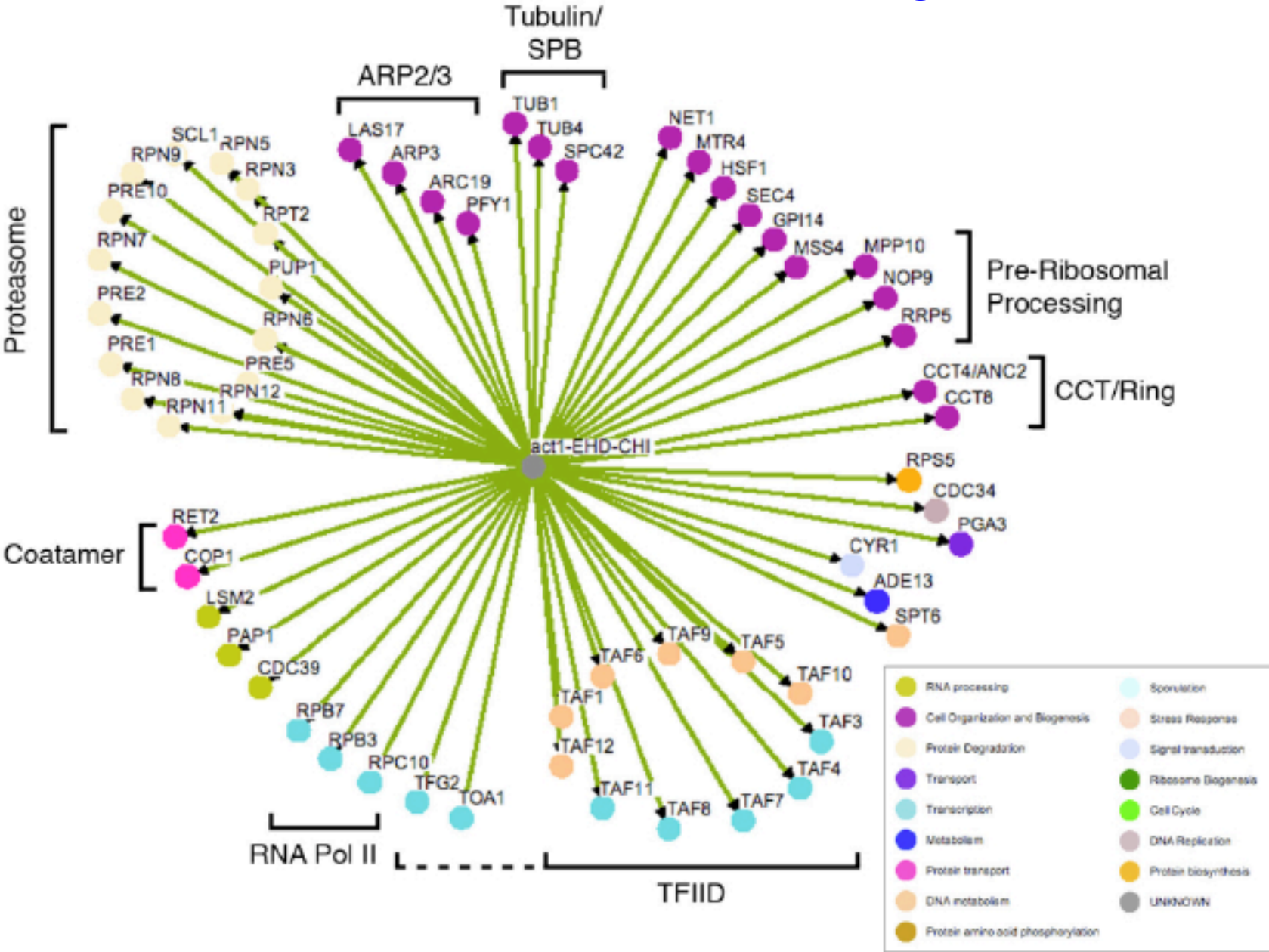
A



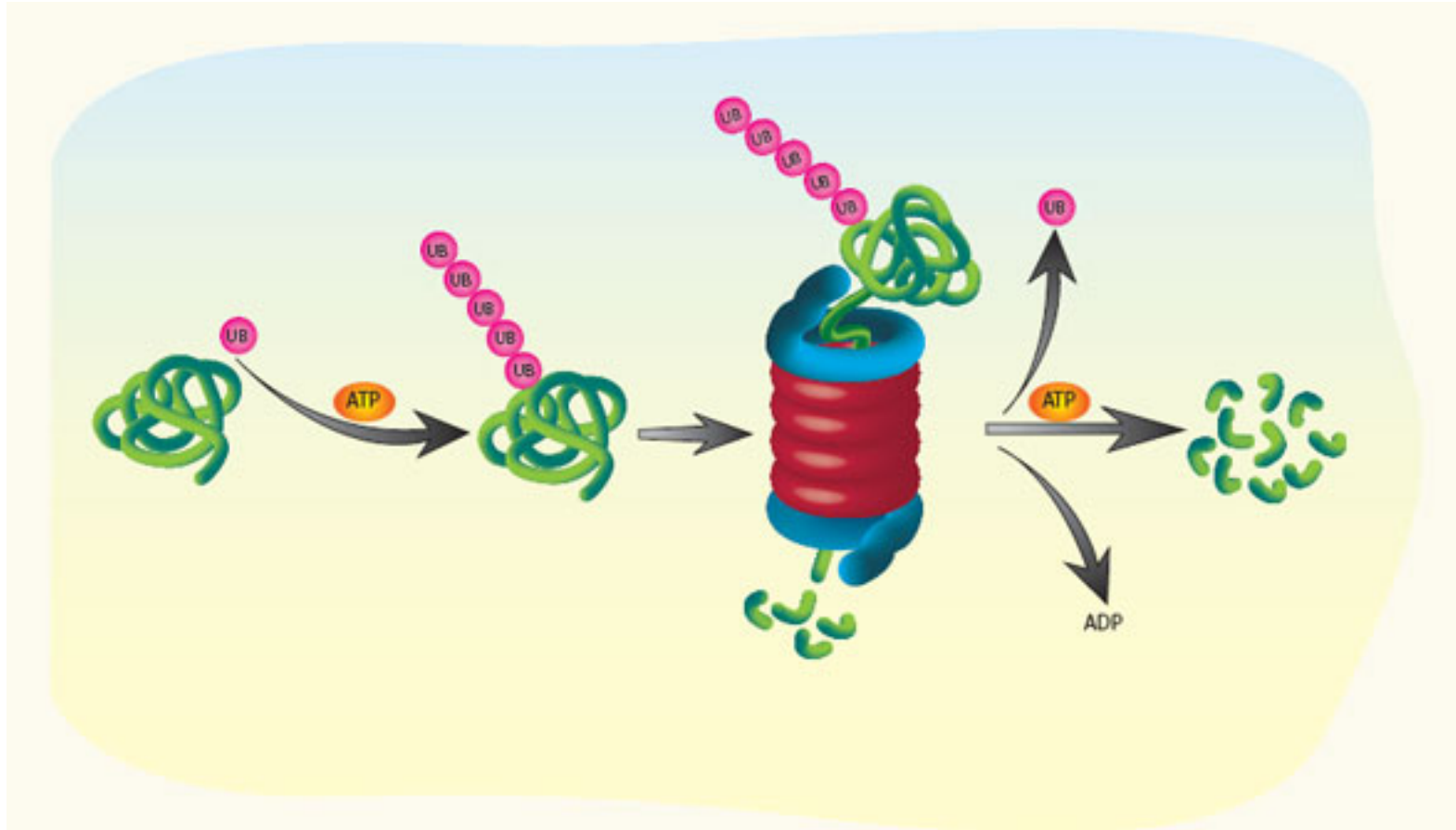
Numbers indicate position of substitutions tested

Colors indicate growth phenotype of substitutions in that region.
Green – wildtype; yellow – conditional;
Red - lethal

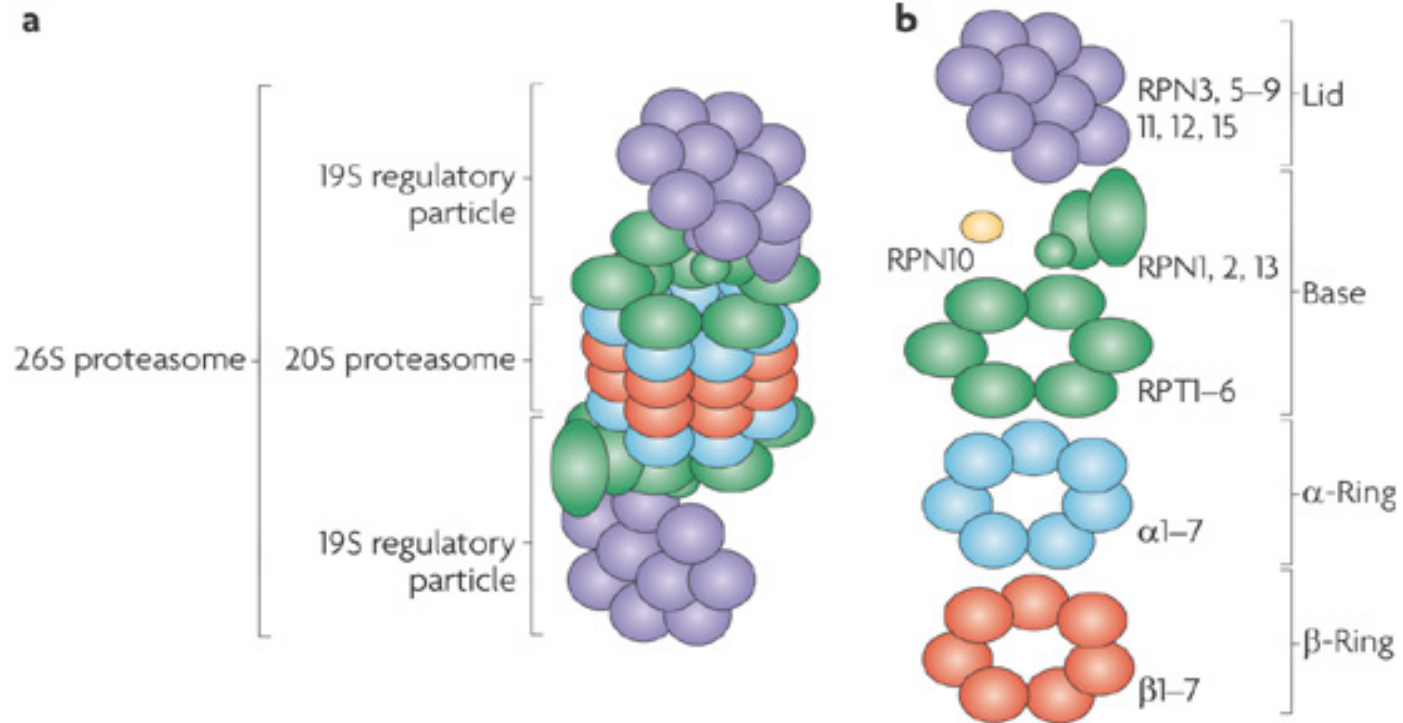
CHI with actin and essential genes



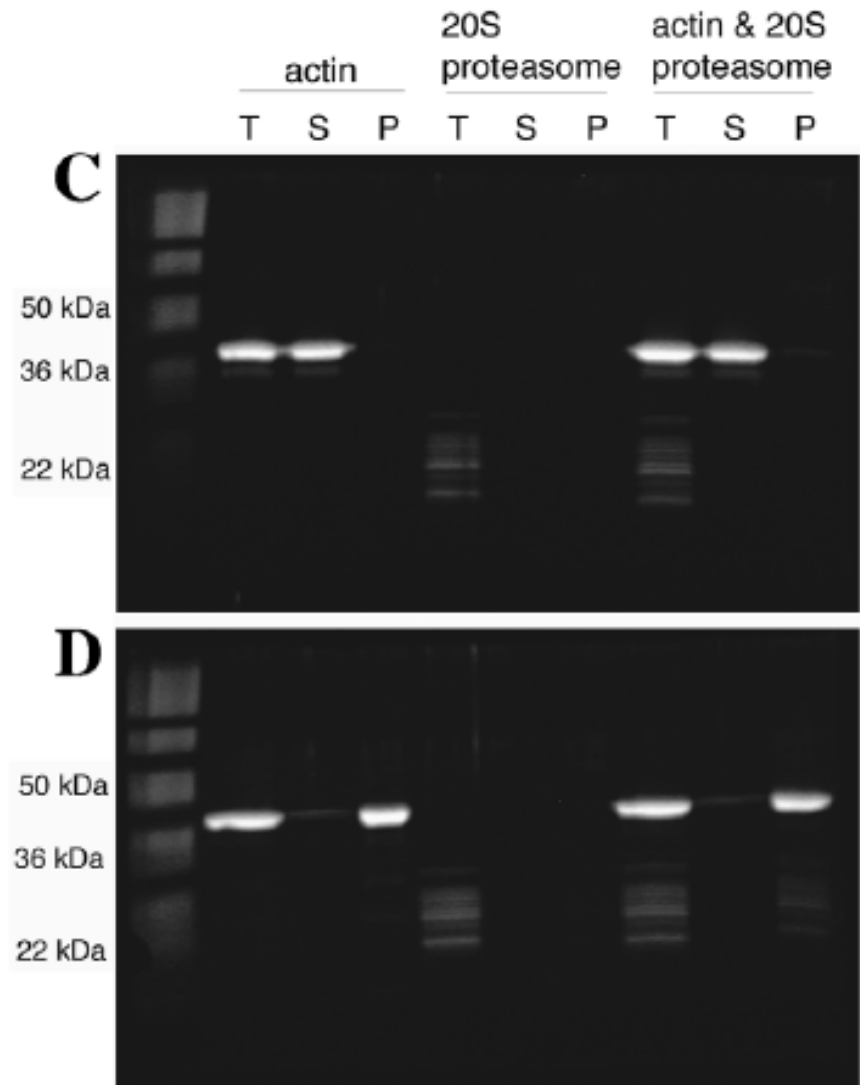
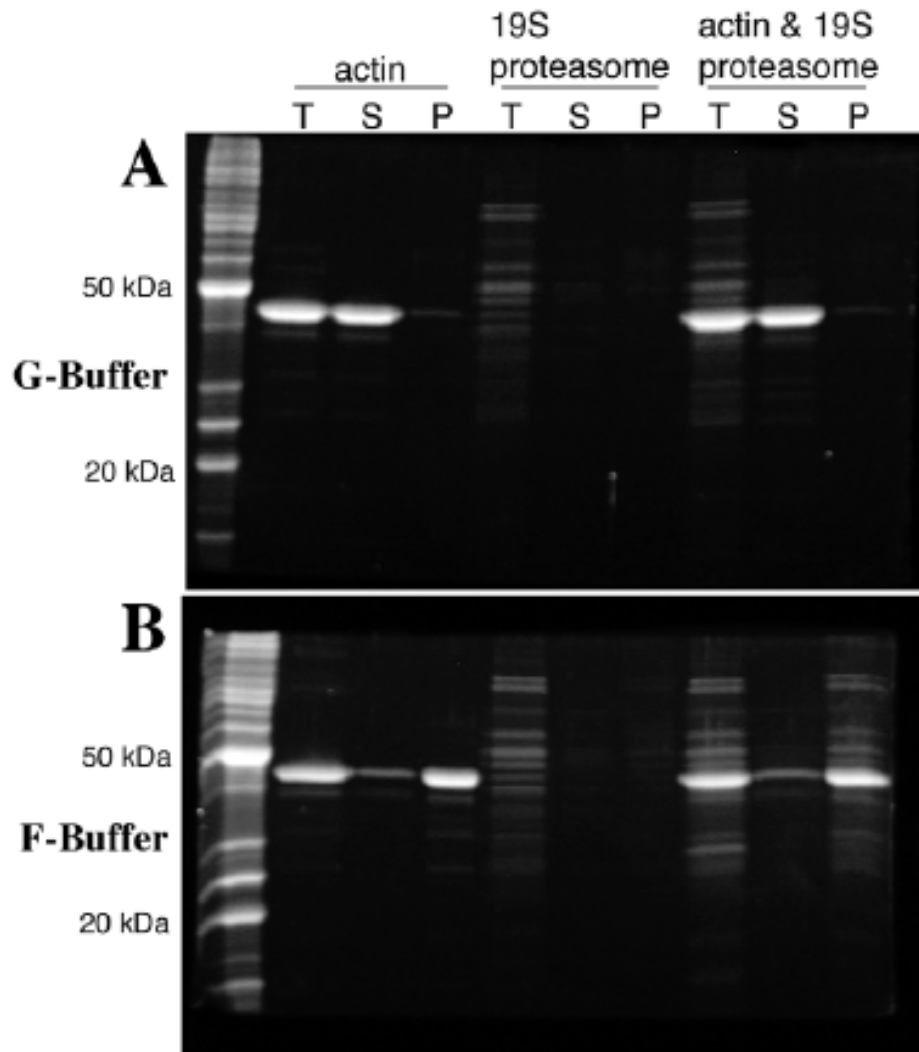
The proteasome and ubiquitin-dependent protein degradation

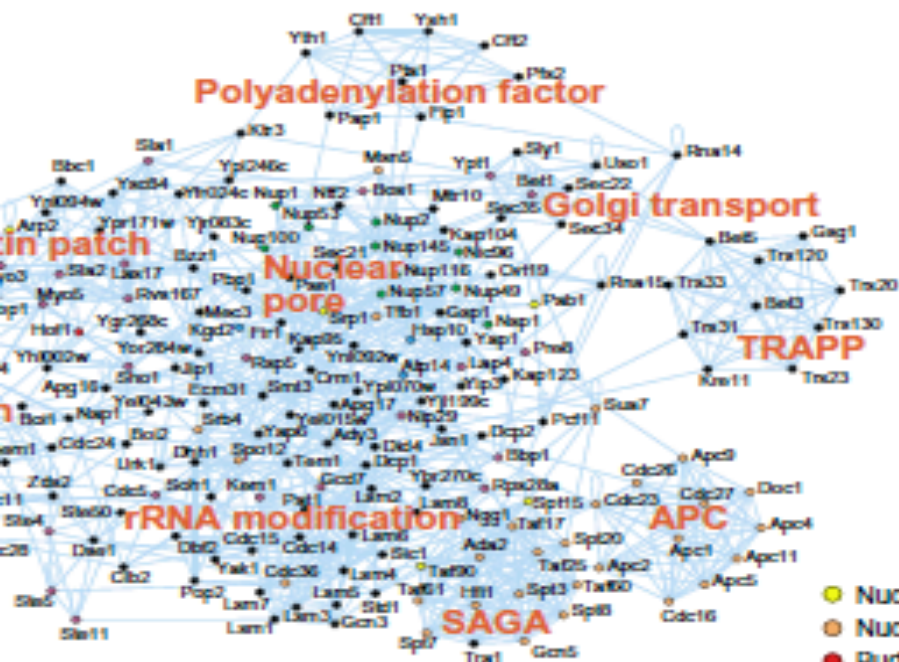


The proteasome consists of two subparticles, 19S and 20S

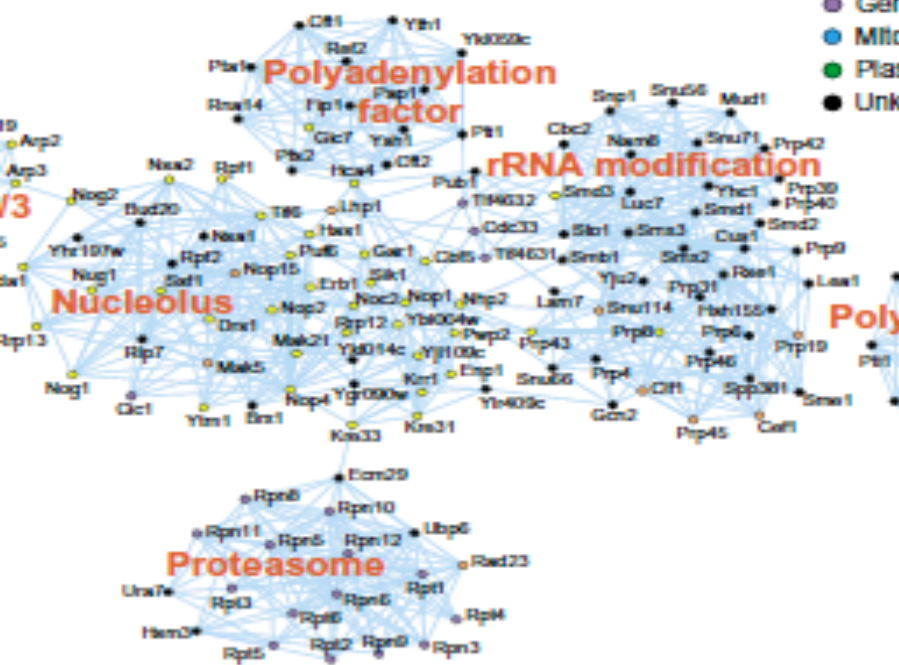
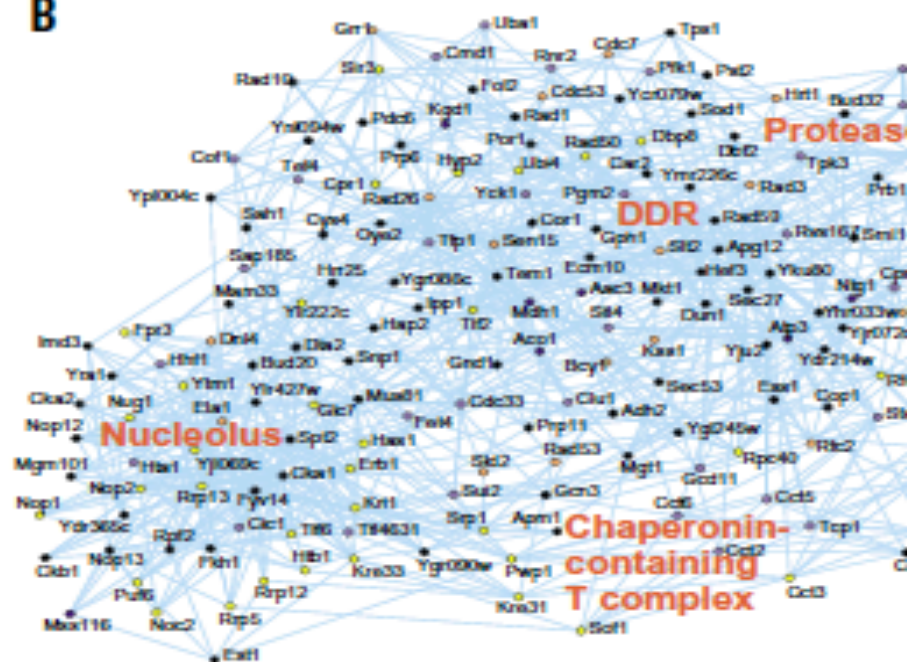


Actin associates with the proteasome





B



D

